

(12) United States Patent

Satoshi

US 9,181,318 B2 (10) **Patent No.:** (45) **Date of Patent:** Nov. 10, 2015

(54) METHOD OF SCREENING A DRUG SUCH AS INSULIN SECRETAGOGUE

(75) Inventor: Inouye Satoshi, Yokohama (JP)

Assignee: JNC Corporation, Tokyo (JP)

(*) Notice: Subject to any disclaimer, the term of this

patent is extended or adjusted under 35

U.S.C. 154(b) by 337 days.

(21) Appl. No.: 12/659,604

(22)Filed: Mar. 15, 2010

(65)**Prior Publication Data**

US 2010/0240038 A1 Sep. 23, 2010

(30)Foreign Application Priority Data

Mar. 16, 2009 (JP) 2009-063279

(2006.01)
(2006.01)
(2006.01)
(2006.01)
(2006.01)
9 (2006.01)
(2006.01)

(52) U.S. Cl.

CPC C07K 14/62 (2013.01); C12N 9/0069 (2013.01); C12Q 1/66 (2013.01); G01N 33/502 (2013.01); G01N 33/74 (2013.01); C07K 2319/61 (2013.01); G01N 2333/62 (2013.01)

(58) Field of Classification Search

See application file for complete search history.

(56)**References Cited**

U.S. PATENT DOCUMENTS

6,194,176 I	B1*	2/2001	Newgard et al	435/69.4
2008/0076156 A	A1*	3/2008	Inouye et al	435/69.7

FOREIGN PATENT DOCUMENTS

EP JP WO WO	1 875 927 2008-99669 WO 99/49019 00/20619 03/088916	1/2008 5/2008 * 9/1999 4/2000 10/2003	 C12N 9/00
WO WO WO	2005/037226 2009/012359 2012/171024	4/2005 1/2009 12/2012	

OTHER PUBLICATIONS

SCORE Result for SEQ ID No. 10; Mar. 2012.*

SCORE Result for SEQ ID No. 8; Mar. 2012.*

Score Report SEQ ID No. 10 Newgard et al downloaded Oct. 19,

Score Report SEQ ID No. 8 Bryan et al downloaded Oct. 19, 2012.* Tang Dissertation entitled Genetic Engineering of Non-Beta-Cells for Regulated Insulin Secretion (Nov. 2003).*

Tannous in Codon-Optimized Gaussia Luciferase cDNA for Mammalian Gene Expression in Culture and in Vivo (Molecular Therapy vol. 11, No. 3, Mar. 2005, pp. 435-443).*

UK Search Report dated Jul. 8, 2010 in Application No. GB1004167.

A. E. Pouli et al., "Insulin Targeting to the Regulated Secretory Pathway after Fusion with Green Fluorescent Protein and Firefly Luciferase", Biochem. J., vol. 331, pp. 669-675, 1998.

Ohara-Imaizumi et al., "Imaging Exocytosis of Single Insulin Secretory Granules with Evanescent Wave Microscopy", The Journal of Biological Chemistry, vol. 277, No. 6, pp. 3805-3808, Feb. 8, 2002. Takahashi et al., "Fusion Pore Dynamics and Insulin Granule Exocytosis in the Pancreatic Islet", Science, vol. 297, pp. 1349-1352,

Inouye et al., "Imagining of Luciferase Secretion from Transformed Chinese Hamster Ovary Cells", Proc. Natl. Acad. Sci., vol. 89, pp. 9584-9587, Oct. 1992

Suzuki et al., "Real-Time Bioluminescence Imagining of a Protein Secretory Pathway in Living Mammalian Cells Using Gaussia Luciferase", FEBS Letters, vol. 581, pp. 4551-4556, 2007.

United Kingdom Office Action issued Mar. 21, 2014 in corresponding United Kingdom Application No. GB1004167.1.

Christian E. Badr et al., "A Highly Sensitive Assay for Monitoring the Secretory Pathway and ER Stress", PLos ONE, vol. 2, Issue 6, Jun. 2007, e571, pp. 1-8.

Office Action issued Mar. 18, 2014 in corresponding Japanese Application No. 2009-063279, with English translation.

Office Action issued Oct. 15, 2013 in corresponding GB Patent Application No. 1309636.7.

Takahiro Suzuki et al., "Video-Rate Bioluminescence Imaging of Matrix Metalloproteinase-2 Secreted from a Migrating Cell", PLoS One, vol. 6, Issue 9, Article No. e25243, Sep. 2011.

Li-Lun Ho et al., "Penta-O-galloyl-β-D-glucose inhibits the invasion of mouse melanoma by suppressing metalloproteinase-9 through down-regulation of activator protein-1", European Journal of Pharmacology, vol. 453, 2002, pp. 149-158.

* cited by examiner

Primary Examiner — Catherine S Hibbert (74) Attorney, Agent, or Firm — Wenderoth, Lind & Ponack, L.L.P.

(57)**ABSTRACT**

The screening method of the present invention is useful for screening drugs such as insulin secretagogues having an insulin secretagogue activity with minimized side effects (hypoglycemia induction, etc.). The transformant in which a polynucleotide encoding the fusion protein used for the screening method is introduced, the screening kit comprising the transformant, etc. are also useful for screening excellent drugs.

5 Claims, 9 Drawing Sheets

FIG. 1

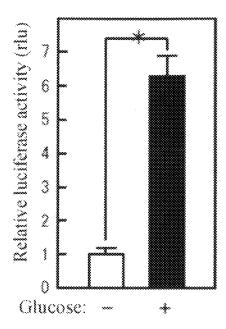


FIG. 2

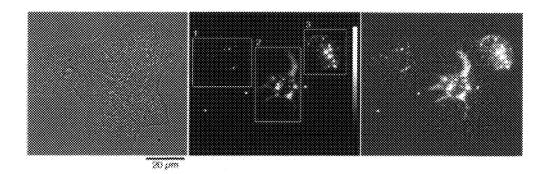
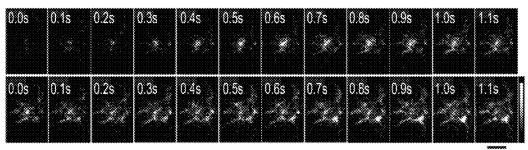


FIG. 3



10 µm

FIG. 4

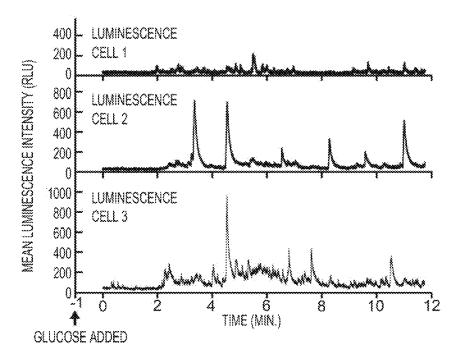


FIG. 5

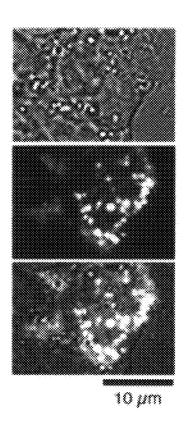


FIG. 6

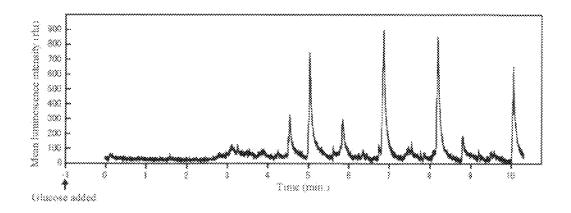


FIG. 7

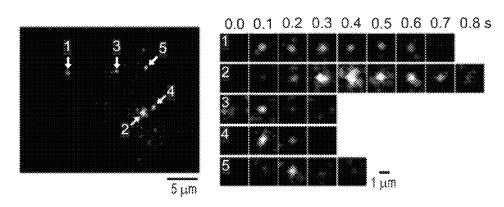


FIG. 8

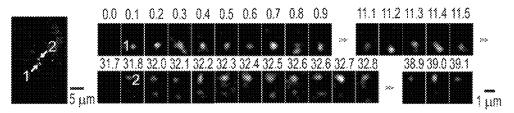


FIG. 9

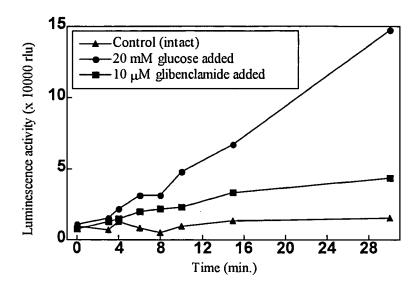


FIG. 10

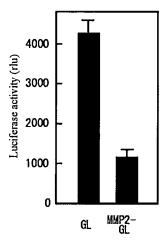


FIG. 11

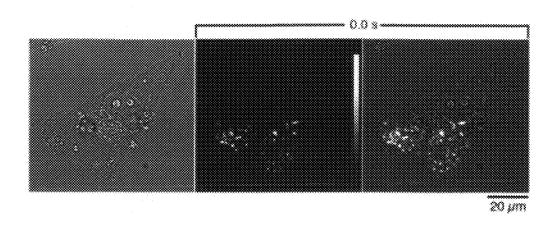


FIG. 12

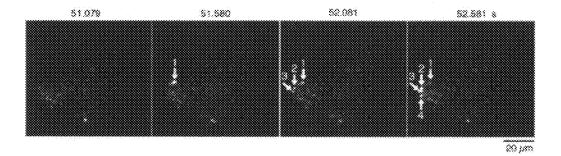


FIG. 13

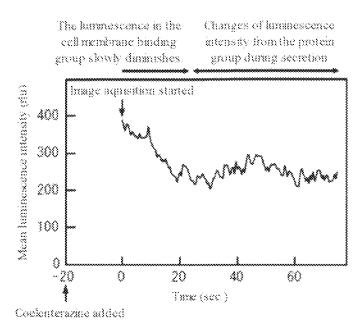


FIG. 14

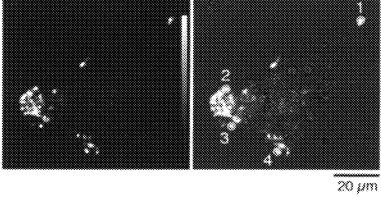


FIG. 15

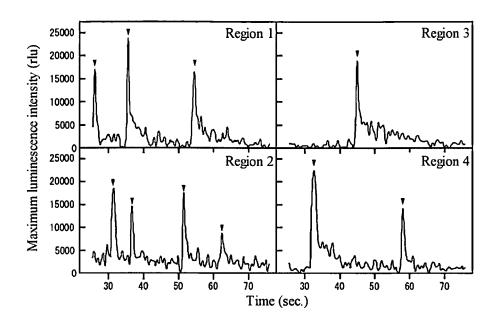


FIG. 16

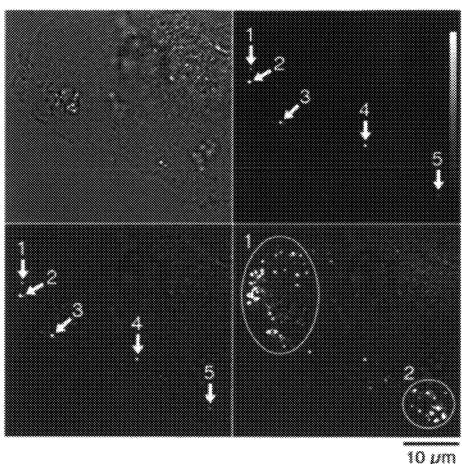
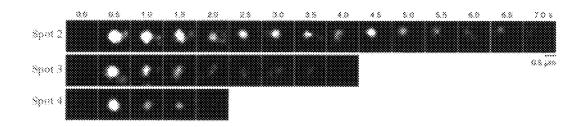


FIG. 17



METHOD OF SCREENING A DRUG SUCH AS INSULIN SECRETAGOGUE

FIELD OF THE INVENTION

The present invention relates to a method of screening a drug such as an insulin secretagogue. More specifically, the present invention relates to a method of screening a drug such as insulin secretagogue using the luminescence imaging method, and so on.

BACKGROUND OF THE INVENTION

Insulin is a hormone that plays an important role in regulating glucose metabolism. Insulin produced from Langerhans' islet β cells in the pancreas is secreted by exocytosis. The secreted insulin acts on cells having insulin receptors to stimulate glucose uptake into cells. Blood sugar levels in the body are maintained in the optimal range by the action of $_{20}$ insulin.

Diabetes mellitus, which is a representative disease associated with insulin, is classified into Type I diabetes mellitus and Type II diabetes mellitus.

In Type I diabetes mellitus, the response to insulin is maintained and blood sugar levels can be controlled by the administration of insulin formulations (e.g., animal insulin preparations extracted from the bovine or swine pancreas; human insulin preparations synthesized by a genetic engineering technique using *Escherichia coli* or a yeast), or by the administration of insulin secretagogues (for example, sulfonylureas (e.g., tolbutamide, glibenclamide, gliclazide, chlorpropamide, tolazamide, acetohexamide, glyclopyramide, glimepiride, glipizide, glybuzole, etc.), repaglinide, senaglinide, nateglinide, mitiglinide, etc.).

The insulin preparations are classified as rapid-acting, regular-acting, intermediate-acting, long-acting, mixed (rapid-acting+intermediate-acting, regular-acting+intermediate-acting), long-acting soluble preparations, and the like, based on time of action. These insulin preparations are appropriately administered depending on the symptoms and conditions of the patient.

As described above, a variety of drugs are known as insulin secretagogues but these drugs involve problems that preprandial hypoglycemia, postprandial hyperglycemia, etc. tend to 45 occur.

Extracellular release (secretion) of a secretory protein is mediated by a mechanism that trafficking vesicles fuse with cell membranes (exocytosis). The total internal reflection fluorescence method (TIRF method) and the fluorescence 50 imaging method (two-photon molecular excitation method) using the excitation of two photons are known as techniques for the visualization of exocytosis.

It has been reported that secretion of the fused protein of enhanced GFP, which is one of fluorescent proteins, with 55 insulin (insulin-EGFP) was observed by the total internal reflection fluorescence method (J. Biol. Chem. 277, 3805-3808 (2002)). More specifically, the phenomenon of exocytosis with the trafficking vesicles containing insulin-EGFP fusion protein was observed from the adhesive part of cells on 60 the side of a glass-bottom dish.

It has been reported that fluorescent dyes were permeated into the spaces where cells are closely adhered to each other, and the secretory vesicles containing insulin fused the cell membranes to form the omega-type structure was observed 65 by the two-photon molecular excitation method (Science, 297, 1349-1352 (2002)).

2

However, both the total internal reflection fluorescence method and the two-photon molecular excitation method have problems that localization of the exocytotic sites on the whole cells cannot be identified and the secreted proteins cannot be quantified, etc.

It has been reported that the exocytotic secretion of luciferase was observed by the bioluminescence imaging method, using Vargula luciferase (Proc. Natl. Acad. Sci., USA, 89, 9584-9587 (1992)).

It has been reported that the process for the secretion of the fused protein (DBHsp-GLase) of the signal peptide sequence (DBHsp) of human DBH (dopamine β -hydroxylase) with Gaussia luciferase (GLase) was observed by the bioluminescence imaging method, (FEBS Letters, 581, 4551-4556 (2007)).

However, no report is known of observation of the exocytotic secretion of functional proteins and functional polypeptides such as hormones, growth factors, etc., by the bioluminescence imaging method.

Matrix metalloproteinase (MMP) is a superfamily of zincdependent endopeptidases and involved in the degradation of the major components of extracellular matrix and connective tissues that inhibit motility of cells.

MMP, especially a gelatinase is known to be associated with metastasis and diffusion of cancers. For example, MMP-2 and MMP-9, which are gelatinases, are known to rise in a particular tumor promoting event. MMP-2 and MMP-9 degrade type IV collagen as the main components of the basal membrane and denatured collagen (gelatin) to induce tumor metastasis. It is also known that disruption of vascular membranes mainly composed of type IV collagen plays an important role in tumor metastasis.

SUMMARY OF THE INVENTION

Under the foregoing circumstances, it has been desired to develop a method of drug screening such as screening of insulin secretagogues having a more potent insulin secretagogue activity with minimized side effects (hypoglycemia induction, etc.) or the like.

It has also been desired to provide a method of drug screening such as screening of more effective tumor metastasis inhibitors (MMP-2 inhibitors, etc.) or the like.

As a result of extensive investigations to solve the problems above, the present inventors have found that by using cells transformed by polynucleotides encoding the fusion proteins of preproinsulins and luciferases, insulin secretion from the cells can be observed by the bioluminescence imaging method thereby to determine localization of the exocytotic sites over the entire cells, quantify the amount of insulin secreted from the cells, and so on.

It has also been found that by using cells transformed by polynucleotides encoding the fusion proteins of pro-MMP-2 and luciferases, secretion of MMP-2 from the cells can be observed by the bioluminescence imaging method thereby to quantify the amount of MMP-2 secreted from the cells, assess diffusion kinetics outside the cells, and so on.

Based on these findings, the inventors have continued further investigations and come to accomplish the present invention

That is, the present invention provides the following features, and so on.

(1) A method of screening a substance regulating insulin secretion from a cell, which comprises using a cell transformed by a polynucleotide encoding a fusion protein of preproinsulin and a luciferase.

- (2) The screening method according to (1) above, wherein the luciferase is a secretory luciferase.
- (3) The screening method according to (2) above, wherein the secretory luciferase is *Gaussia* luciferase.
- (4) The screening method according to (3) above, wherein *Gaussia* luciferase is a protein of any one of (a) through (d) below:
- (a) a protein comprising the amino acid sequence of SEQ ID NO: 8;
- (b) a protein consisting of an amino acid sequence in which at least one amino acid residue is deleted, substituted, inserted and/or added in the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity;
- (c) a protein consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity; and,
- (d) a protein consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 7, and having a luciferase activity.
- (5) The screening method according to (4) above, wherein *Gaussia* luciferase is a protein of any one of (a) through (d) below:
- (a) a protein consisting of the amino acid sequence of SEQ ID NO: 8;
- (b) a protein consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity;
- (c) a protein consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity; and,
- (d) a protein consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 7, and having a luciferase activity.
- (6) The screening method according to any one of (1) to (5) above, wherein preproinsulin is a polypeptide of any one of (e) through (h) below:
- (e) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 10;
- (f) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 10, and having an insulin activity;
- (g) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 10, and having an insulin activity; and,
- (h) a polypeptide comprising a polypeptide consisting of 55 an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 9, and having an insulin activity.
- (7) The screening method according to (6) above, wherein preproinsulin consists of a signal peptide of preproinsulin and the polypeptide of any one of (i) through (l) below:
- (i) a polypeptide consisting of the amino acid sequence of SEO ID NO: 4:
- (j) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, inser-

4

tion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 4, and having an insulin activity;

- (k) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 4, and having an insulin activity; and.
- (l) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 3, and having an insulin activity.
- (8) The screening method according to (7) above, wherein the signal peptide of preproinsulin is a polypeptide consisting of the amino acid sequence of SEQ ID NO: 6.
 - (9) The screening method according to (1) through (8) above, wherein the fusion protein is a polypeptide of any one of (m) through (p) below:
 - (m) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 12;
 - (n) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 12, and having an insulin activity;
 - (o) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 12, and having an insulin activity; and.
 - (p) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 11, and having an insulin activity.
 - (10) The screening method according to (1) through (9) above, which further comprises the step of detecting luminescence using a CCD camera or a photon counting camera.
 - (11) A polynucleotide encoding a fusion protein of preproinsulin and a luciferase.
 - (12) The polynucleotide according to (11) above, wherein the luciferase is a secretory luciferase.
- (13) The polynucleotide according to (12) above, wherein the secretory luciferase is *Gaussia* luciferase.
- (14) The polynucleotide according to (13) above, wherein *Gaussia* luciferase is a protein of any one of (a) through (d) below:
- (a) a protein comprising the amino acid sequence of SEQ50 ID NO: 8;
 - (b) a protein consisting of an amino acid sequence in which at least one amino acid residue is deleted, substituted, inserted and/or added in the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity;
 - (c) a protein consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity; and,
- (d) a protein consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 7, and having a luciferase activity.
- (15) The polynucleotide according to (14) above, wherein *Gaussia* luciferase is a protein according to any one of (a) through (d) below:
 - (a) a protein consisting of the amino acid sequence of SEQ ID NO: 8;

- (b) a protein consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity;
- (c) a protein consisting of an amino acid sequence having at 5 least 90% identity to the amino acid sequence of SEO ID NO: 8, and having a luciferase activity; and,
- (d) a protein consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 7, and having a luciferase activity.
- (16) The polynucleotide according to any one of (1) through (15) above, wherein preproinsulin is a polypeptide 15 according to any one of (e) through (h) below:
- (e) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 10;
- (f) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, inser-20 tion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 10, and having an insulin activity;
- (g) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the 25 amino acid sequence of SEQ ID NO: 10, and having an insulin activity; and,
- (h) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 9, and having an insulin activity.
- (17) The polynucleotide according to (16) above, wherein preproinsulin consists of a signal peptide of preproinsulin and the polypeptide according to any one of (i) through (l) below:
- (i) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 4;
- (j) a polypeptide comprising a polypeptide consisting of an 40 amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 4, and having an insulin activity;
- (k) a polypeptide comprising a polypeptide consisting of 45 luciferase is a secretory luciferase. an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 4, and having an insulin activity; and,
- (1) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that 50 hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 3, and having an insulin activity.
- (18) The polynucleotide according to (17) above, wherein 55 the signal peptide of preproinsulin is a polypeptide consisting of the amino acid sequence of SEQ ID NO: 6.
- (19) The polynucleotide according to any one of (11) through (18) above, wherein the fusion protein is a polypeptide according to any one of (m) through (p) below:
- (m) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 12;
- (n) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues 65 in the amino acid sequence of SEQ ID NO: 12, and having an insulin activity;

6

- (o) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 12, and having an insulin activity; and,
- (p) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 11, and having an insulin activity.
- (20) A recombinant vector comprising the polynucleotide according to any one of (11) through (19) above.
- (21) A transformant wherein the recombinant vector according to (20) above is introduced.
- (22) The transformant according to (21) above, which is derived from a cell line.
- (23) The transformant according to (22) above, which is derived from a mammal.
- (24) The transformant according to any one of (21) to (23) above, which is derived from a pancreatic β cell.
- (25) A method of observing the extracellular secretion of insulin, which comprises using the transformant according to any one of (21) to (24) above.
- (26) A method of identifying the localization of exocytotic site of insulin in a cell, which comprises using the transformant according to any one of (21) to (24) above.
- (27) A method of quantifying insulin secreted extracellularly, which comprises using the transformant according to any one of (21) to (24) above.
- (28) A method of determining the frequency of insulin secretion, which comprises using the transformant according to any one of (21) to (24) above.
- (29) A method of observing the diffusion dynamics of 35 insulin secreted extracellularly, which comprises using the transformant according to any one of (21) to (24) above.
 - (30) A kit comprising the transformant according to any one of (21) to (24) above.
 - (31) The kit according to (30) above, which is a kit used for screening a drug.
 - (32) The kit according to (30) or (31) above, further comprising a luciferin.
 - (33) A fusion protein of preproinsulin and a luciferase.
 - (34) The protein according to (33) above, wherein the
 - (35) The protein according to (34) above, wherein the secretory luciferase is Gaussia luciferase.
 - (36) The protein according to (35) above, wherein Gaussia luciferase is a protein according to any one of (a) through (d) below.
 - (a) a protein comprising the amino acid sequence of SEQ ID NO: 8;
 - (b) a protein consisting of an amino acid sequence in which at least one amino acid residue is deleted, substituted, inserted and/or added in the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity;
 - (c) a protein consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity; and,
 - (d) a protein consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 7, and having a luciferase activity.
 - (37) The protein according to (36) above, wherein Gaussia luciferase is a protein according to any one of (a) through (d) below.

- (a) a protein consisting of the amino acid sequence of SEQ
- (b) a protein consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of 5 SEQ ID NO: 8, and having a luciferase activity;
- (c) a protein consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity; and,
- (d) a protein consisting of an amino acid sequence encoded 10 by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 7, and having a luciferase activity.
- (38) The protein according to any one of (33) through (37) 15 above, wherein preproinsulin is a polypeptide according to any one of (e) through (h) below.
- (e) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 10;
- (f) a polypeptide comprising a polypeptide consisting of an 20 wherein the luciferase is a secretory luciferase. amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 10, and having an insulin activity;
- (g) a polypeptide comprising a polypeptide consisting of 25 an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 10, and having an insulin activity; and,
- (h) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that 30 hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 9, and having an insulin activity.
- (39) The protein according to (38) above, wherein prepro- 35 insulin consists of a signal peptide of preproinsulin and the polypeptide according to any one of (i) through (l) below:
- (i) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 4;
- (j) a polypeptide comprising a polypeptide consisting of an 40 amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 4, and having an insulin activity;
- (k) a polypeptide comprising a polypeptide consisting of 45 an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 4, and having an insulin activity; and,
- (1) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that 50 hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 3, and having an insulin activity.
- (40) The protein according to (39) above, wherein the 55 signal peptide of preproinsulin is a polypeptide consisting of the amino acid sequence of SEQ ID NO: 6.
- (41) The protein according to any one of (33) through (40) above, comprising the polypeptide according to any one of (m) through (p) below:
- (m) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 12;
- (n) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues 65 in the amino acid sequence of SEQ ID NO: 12, and having an insulin activity;

- (o) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 12, and having an insulin activity; and,
- (p) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 11, and having an insulin activity.

The present invention further provides the following features, and so on.

- (1B) A method of screening a drug such as a substance inhibiting the secretion and/or activity of MMP-2, a cancer metastasis inhibitor, etc., which comprises using a cell transformed by a polynucleotide encoding a fusion protein of pro-MMP-2 and a luciferase.
- (2B) The screening method according to (1B) above,
- (3B) The screening method according to (2B) above, wherein the secretory luciferase is Gaussia luciferase.
- (4B) The screening method according to (3B) above, wherein Gaussia luciferase is a protein of any one of (a) through (d) below:
- (a) a protein comprising the amino acid sequence of SEQ ID NO: 8;
- (b) a protein consisting of an amino acid sequence in which at least one amino acid residue is deleted, substituted, inserted and/or added in the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity;
- (c) a protein consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity; and,
- (d) a protein consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 7, and having a luciferase activity.
- (5B) The screening method according to (4B) above. wherein Gaussia luciferase is a protein of any one of (a) through (d) below:
- (a) a protein consisting of the amino acid sequence of SEQ ID NO: 8:
- (b) a protein consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity;
- (c) a protein consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity; and,
- (d) a protein consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 7, and having a luciferase activity.
- (6B) The screening method according to any one of (1B) to (5B) above, wherein pro-MMP-2 is a polypeptide of any one 60 of (e) through (h) below:
 - (e) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 18;
 - (f) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 18, and having a MMP-2 activity;

- (g) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 18, and having a MMP-2 activity; and,
- (h) a polypeptide comprising a polypeptide consisting of 5 an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 17, and having a MMP-2 activity.
- (7B) The screening method according to (6B) above, wherein pro-MMP-2 consists of a signal peptide of pro-MMP-2 and the polypeptide of any one of (i) through (l) below:
- (i) a polypeptide consisting of the amino acid sequence of 15 SEQ ID NO: 14;
- (j) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 14, and having an 20 ID NO: 8; insulin activity; wherein G of (a) throw the original of the comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 14, and having an 20 ID NO: 8; insulin activity;
- (k) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 14, and having an insulin activity; and,
- (l) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 13, and having an 30 insulin activity.
- (8B) The screening method according to (7B) above, wherein the signal peptide of pro-MMP-2 is a polypeptide consisting of the amino acid sequence of SEQ ID NO: 16.
- (9B) The screening method according to (1B) through (8B) 35 above, wherein the fusion protein is a polypeptide of any one of (m) through (p) below:
- (m) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 20;
- (n) a polypeptide comprising a polypeptide consisting of 40 an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 20, and having an insulin activity;
- (o) a polypeptide comprising a polypeptide consisting of 45 an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 20, and having an insulin activity; and,
- (p) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that 50 hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 19, and having an insulin activity.
- (10B) The screening method according to (1B) through 55 (9B) above, which further comprises the step of detecting luminescence using a CCD camera or a photon counting camera.
- (11B) A polynucleotide encoding a fusion protein of pro-MMP-2 and a luciferase.
- (12B) The polynucleotide according to (11B) above, wherein the luciferase is a secretory luciferase.
- (13B) The polynucleotide according to (12B) above, wherein the secretory luciferase is *Gaussia* luciferase.
- (14B) The polynucleotide according to (13B) above, 65 wherein *Gaussia* luciferase is a protein of any one of (a) through (d) below:

10

- (a) a protein comprising the amino acid sequence of SEQ ID NO: 8;
- (b) a protein consisting of an amino acid sequence in which at least one amino acid residue is deleted, substituted, inserted and/or added in the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity;
- (c) a protein consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity; and,
- (d) a protein consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 7, and having a luciferase activity.
- (15B) The polynucleotide according to (14B) above, wherein *Gaussia* luciferase is a protein according to any one of (a) through (d) below:
- (a) a protein consisting of the amino acid sequence of SEQ ID NO: 8:
- (b) a protein consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity;
- (c) a protein consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity; and,
- (d) a protein consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 7, and having a luciferase activity.
- (16B) The polynucleotide according to any one of (1B) through (15B) above, wherein pro-MMP-2 is a polypeptide according to any one of (e) through (h) below:
- (e) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 18;
- (f) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 18, and having an insulin activity;
- (g) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 18, and having an insulin activity; and,
- (h) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 17, and having an insulin activity.
- (17B) The polynucleotide according to (16B) above, wherein pro-MMP-2 consists of a signal peptide of pro-MMP-2 and the polypeptide according to any one of (i) through (l) below:
- (i) a polypeptide consisting of the amino acid sequence of SEO ID NO: 14;
- (j) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 14, and having an insulin activity;
 - (k) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 14, and having an insulin activity; and,

- (I) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 13, and having an insulin activity.
- (18B) The polynucleotide according to (17B) above, wherein the signal peptide of pro-MMP-2 is a polypeptide consisting of the amino acid sequence of SEQ ID NO: 16.
- (19B) The polynucleotide according to any one of (11B) through (17B) above, wherein the fusion protein is a polypeptide according to any one of (m) through (p) below:
- (m) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 20;
- (n) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 20, and having an insulin activity;
- (o) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 20, and having an insulin activity; and,
- (p) a polypeptide comprising a polypeptide consisting of 25 an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 19, and having an insulin activity.
- (20B) A recombinant vector comprising the polynucleotide according to any one of (11B) through (19B) above.
- (21B) A transformant wherein the recombinant vector according to (20B) above is introduced.
- (22B) The transformant according to (21B) above, which is derived from a cell line.
- $(23B)\, The transformant according to (22B) above, which is derived from a mammal.$
- (24B) The transformant according to any one of (21B) to $_{40}$ (23B) above, which is derived from a cancer cell.
- (25B) A method of observing the extracellular secretion of MMP-2, which comprises using the transformant according to any one of (21B) to (24B) above.
- (26B) A method of identifying the localization of exocytotic site of MMP-2 in a cell, which comprises using the transformant according to any one of (21B) to (24B) above.
- (27B) A method of quantifying MMP-2 secreted extracellularly, which comprises using the transformant according to any one of (21B) to (24B) above.
- (28B) A method of determining the frequency of insulin secretion, which comprises using the transformant according to any one of (21B) to (24B) above.
- (29B) A method of observing the diffusion dynamics of MAP-2 secreted extracellularly, which comprises using the 55 transformant according to any one of (21B) to (24B) above.
- (30B) A kit comprising the transformant according to any one of (21B) to (24B) above.
- (31B) The kit according to (30B) above, which is a kit used for screening a drug.
- (32B) The kit according to (30B) or (31B) above, further comprising a luciferin.
 - (33B) A fusion protein of pro-MMP-2 and a luciferase.
- (34B) The protein according to (33B) above, wherein the luciferase is a secretory luciferase.
- (35B) The protein according to (34B) above, wherein the secretory luciferase is *Gaussia* luciferase.

12

- (36B) The protein according to (35B) above, wherein *Gaussia* luciferase is a protein according to any one of (a) through (d) below.
- (a) a protein comprising the amino acid sequence of SEQ ID NO: 8;
- (b) a protein consisting of an amino acid sequence in which at least one amino acid residue is deleted, substituted, inserted and/or added in the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity;
- (c) a protein consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity; and,
- (d) a protein consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 7, and having a luciferase activity.
- (37B) The protein according to (36B) above, wherein 20 *Gaussia* luciferase is a protein according to any one of (a) through (d) below.
 - (a) a protein consisting of the amino acid sequence of SEQ ID NO: 8;
 - (b) a protein consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity;
 - (c) a protein consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 8, and having a luciferase activity; and,
- (d) a protein consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 7, and having a luciferase activity.
 - (38B) The protein according to any one of (33B) through (37B) above, wherein pro-MMP-2 is a polypeptide according to any one of (e) through (h) below.
 - (e) a polypeptide consisting of the amino acid sequence of SEO ID NO: 18:
 - (f) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 18, and having a MMP-2 activity;
 - (g) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 18, and having a MMP-2 activity; and.
 - (h) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 18, and having a MMP-2 activity.
- (39B) The protein according to (38B) above, wherein pro-MMP-2 consists of a signal peptide of pro-MMP-2 and the polypeptide according to any one of (i) through (l) below:
 - (i) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 14;
 - (j) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 14, and having a MMP-2 activity;

(k) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 14, and having a MMP-2 activity; and.

(l) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 13, and having a MMP-2 activity.

(40B) The protein according to (39B) above, wherein the signal peptide of pro-MMP-2 is a polypeptide consisting of the amino acid sequence of SEQ ID NO: 16.

(41B) The protein according to any one of (33B) through (40B) above, comprising the polypeptide according to any one of (m) through (p) below:

(m) a polypeptide consisting of the amino acid sequence of SEO ID NO: 20;

(n) a polypeptide comprising a polypeptide consisting of 20 an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 20, and having a MMP-2 activity;

(o) a polypeptide comprising a polypeptide consisting of 25 an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 20, and having a MMP-2 activity; and,

(p) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that ³⁰ hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 19, and having a MMP-2 activity.

According to the present invention, the method of drug screening such as screening of insulin secretagogues having a more potent insulin secretagogue activity with minimized side effects (hypoglycemia induction, etc.), etc. can be provided. Furthermore, the transformants in which the polynucleotide encoding the fusion protein used in the screening method is introduced, the screening kit comprising the transformants, etc. can be provided.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 shows the comparison of activities of the insulin-GLase fusion protein secreted outside cells by the addition or without addition (-, +) to the transformant derived from the MIN6 cell line obtained in EXAMPLE 1-2.

FIG. 2 shows the luminescence images of insulin-GLase 50 fusion protein secreted by stimulation of glucose added to the transformant derived from the MIN6 cell line obtained in EXAMPLE 1-3. Left, bright-field image; center, luminescence signal image; right, image obtained by superimposing the luminescent image on the bright-field image.

FIG. 3 shows the images of luminescence signals acquired at 100 msec, indicating time-lapse changes of the insulin-GLase fusion protein secreted in a periodic pulsatile manner by stimulation of glucose added to the transformant derived from the MIN6 cell line obtained in EXAMPLE 1-3.

FIG. 4 is graphs formed by quantifying time-lapse changes of the insulin-GLase fusion protein secreted by stimulation of glucose added, in the cell regions 1 to 3 designated in FIG. 2.

FIG. 5 shows the luminescence images showing the distribution of the insulin-GLase fusion protein secreted by stimulation of glucose added to the transformant derived from the MIN6 cell line obtained in EXAMPLE 1-3. Upper: bright-

14

field image, center: luminescence signal image, lower: image superimposed on the bright-field image.

FIG. 6 shows the graphs formed by quantifying time-lapse changes of the insulin-GLase fusion protein secreted in a periodic pulsatile manner by stimulation of glucose added, in the image regions of FIG. 5, from the mean luminescence intensity.

FIG. 7 shows the distribution of the insulin-GLase fusion protein by exocytosis, not secreted in a periodic pulsatile manner, by stimulation of glucose added to the transformant derived from the MIN6 cell line obtained in EXAMPLE 1-3 and the images showing time-lapse changes of the luminescence signal at each luminescence spot for a short period of time (within a second).

FIG. 8 shows the distribution of the insulin-GLase fusion protein by exocytosis, not secreted in a periodic pulsatile manner, by stimulation of glucose added to the transformant derived from the MIN6 cell line obtained in EXAMPLE 1-3 and the images showing time-lapse changes of the luminescence signal at each luminescence spot for a long period of time (more than a second).

FIG. 9 shows the graphs showing a drug efficacy evaluation of glibenclamide as a hypoglycemic agent using the insulin-GLase-expressed MIN6 cells.

FIG. 10 shows the graph comparing the activities of the hMMP-2-GLase fusion protein (MMP2-GL) secreted from the transformant derived from the HeLa cell line obtained in EMBODIMENT 3-2 and

FIG. 11 shows the luminescence images showing the distribution of the hMMP-2-GLase fusion protein secreted from the transformant derived from the HeLa cell line obtained in EMBODIMENT 3-2 in cell membrane. Left, bright-field image, center: luminescence signal image, right: image superimposed on the bright-field image.

FIG. 12 shows the luminescence images showing the timelapse distribution of the hMMP-2-GLase fusion protein from the transformant derived from the HeLa cell line obtained in EMBODIMENT 3-2 in cell membrane.

FIG. 13 shows the graph showing changes of the mean luminescence intensity of the hMMP-2-GLase fusion protein from the transformant derived from the HeLa cell line obtained in EXAMPLE 3-2, with the lapse of time.

FIG. 14 shows the luminescence images of the hMMP-2-GLase fusion protein from the transformant derived from the HeLa cell line obtained in Example 3-2 in cell membrane. Left: luminescence signal image, right: image superimposed on the bright-field image.

FIG. 15 is graphs formed by quantifying time-lapse changes of the hMMP-2-GLase fusion protein in the luminescence spot regions 1 to 4 designated in FIG. 14, wherein the arrow designates potent secretory luminescence signals.

FIG. 16 shows the luminescence images of the hMMP-2-GLase fusion protein from the transformant derived from the HeLa cell line obtained in Example 3-2, at the leading edge. Upper left: bright-field images, upper right: luminescence signal images, lower left: luminescence images superimposed on the bright-field images, lower right: images showing the luminescence spot region at the leading edge appeared for 100 seconds.

FIG. 17 shows time-lapse changes of the luminescence images of the hMMP-2-GLase fusion protein in the luminescence spots 2, 3 and 4 designated in FIG. 16.

DETAILED DESCRIPTION OF THE INVENTION

1. Drug Screening Method of the Invention

The drug screening method of the invention is directed to a method of drug screening including screening of a substance

that promotes (or regulates) insulin secretion from cells, a substance that inhibits the secretion and/or activity of MMP-2, a cancer metastasis inhibitor, etc., using the cells transformed by the polynucleotide encoding the fusion protein of a secretory protein such as preproinsulin or MMP-2 and a 5 luciferase.

More specifically, the drug screening method of the present invention is directed to a method of screening a substance that promotes (or regulates) insulin secretion from cells, which comprises observing the insulin secretion from cells by the bioluminescence imaging method, and comparing the insulin levels secreted in the cases where a test compound is added or not added, using the cell transformed by the polynucleotide encoding the fusion protein of, e.g., preproinsulin and the luciferase.

The drug screening method of the invention is also directed to a method of drug screening including screening of a substance that inhibits the secretion and/or activity of MMP-2, a cancer metastasis inhibitor, etc., by comparing the secretion of MMP-2, etc., which comprises comparing the secretion of 20 luciferin-luciferase reaction with luciferin as a luminescence MMP-2, etc. in the cases where a test compound is added or not added, using the cell transformed by the polynucleotide encoding the fusion protein of, e.g., pro-MMP-2 and the luciferase.

Hereinafter, the present invention is described in detail 25 with reference to the embodiments.

The drug screening method of the invention includes, for example, Screening method 1, Screening method 2, and so

Screening Method 1:

The method of screening a substance that regulates insulin secretion from cells, which comprises Steps (a) through (d) below:

Step (a): the step of culturing a cell transformed by the polynucleotide encoding the fusion protein of preproinsulin 35 and the luciferase in a medium in the presence or absence of a test compound;

Step (b): the step of expressing the fusion protein of the present invention to secrete the fusion protein of the invention into a luciferin-containing medium (outside the cells).

Step (c): the step of observing the fusion protein secreted extracellularly by luminescence imaging; and,

Step (d): the step of comparing the expression of the fusion protein in the presence of or in the absence of a test compound.

These steps are explained below in detail.

Step (a)

(1) Luciferase

The luciferase used in the present invention refers to an enzyme which catalyzes the reaction that luciferin as a lumi- 50 nescence substrate is oxidized in the presence of oxygen to form oxyluciferin. This oxidization reaction by the oxygen of luciferin which is catalyzed by a luciferase is called a luciferin-luciferase reaction. The oxyluciferin formed by the luciferin-luciferase reaction is formed in an excited state, and 55 emission of light occurs during a transition to the ground

The luciferase used in the present invention is preferably a secretory luciferase, more preferably, a coelenterazine-type luciferase, and particularly preferably Gaussia luciferase. 60 Examples of the coelenterazine-type luciferase include Gaussia luciferase, Renilla (Renilla reniformis) luciferase, Pleuromamma luciferase, Metridia luciferase, Oplophorus (Oplophorus grachlorostris) luciferase, and the like.

The secretory luciferase refers to a luciferase having a 65 secretory signal peptide, which can be secreted extracellularly.

16

As used herein, where the luciferase is a secretory luciferase, the luciferase may have a signal peptide or may lack a signal peptide.

Gaussia luciferase means a luciferase derived from Gaussia princeps. As used herein, Gaussia luciferase may have a signal peptide or may lack a signal peptide. In other words, Gaussia luciferase lacking a signal peptide is included in Gaussia luciferase.

Gaussia luciferase used in the present invention includes a protein consisting of the amino acid sequence of SEQ ID NO: 2, a protein consisting of the amino acid sequence of SEQ ID NO: 8, a protein having substantially the same activity or function as the protein consisting of the amino acid sequence of SEQ ID NO: 2, and a protein having substantially the same activity or function as the protein consisting of the amino acid sequence of SEQ ID NO: 8.

The term "substantially the same activity or function" is used to mean that, for example:

- (i) the protein described above functions to effect the
- (ii) the maximum luminescence intensity (Imax) of the luminescence caused by the luciferin-luciferase reaction with the protein above is \(^{1}\sqrt{4}\) or more, preferably \(^{1}\sqrt{3}\) or more, more preferably ½ or more and most preferably 1/1.5 or more, than the maximum luminescence intensity of the luminescence caused by the luciferin-luciferase reaction with the amino acid sequence of SEQ ID NO: 8;

(iii) the half-life period of the luminescence caused by the luciferin-luciferase reaction with the protein above is 4 times or less, preferably 3 times or less, more preferably 2 times or less and most preferably 1.5 times or less, than the half-life period of the luminescence caused by the luciferin-luciferase reaction with the amino acid sequence of SEQ ID NO: 8; and so on. The substantially the same activity or function is sometimes also referred to as the "luciferase activity." The luminescence activity or luminescence pattern described above may be measured by the methods described in, e.g., Methods in Enzymology 326, 165-174 (2000), etc. Specifically, 40 luciferin is added to the protein above in the presence of oxygen to initiate the luminescence reaction, and the luminescence activity or luminescence pattern may be measured using a luminescence measurement device, for example, TD-4000 (manufactured by Labo Science), Berthold 960 (manufactured by Berthold, Inc.), etc.

More specifically, the Gaussia luciferase used in the present invention includes (a) a protein comprising the amino acid sequence of SEQ ID NO: 8; (b) a protein consisting of an amino acid sequence in which at least one amino acid residue is deleted, substituted, inserted and/or added in the amino acid sequence of SEQ ID NO: 8, and having the luciferase activity; (c) a protein consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 8, and having the luciferase activity; (d) a protein consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 7, and having the luciferase activity; (a') a protein consisting of the amino acid sequence of SEQ ID NO: 2; (b') a protein consisting of an amino acid sequence in which at least one amino acid residue is deleted, substituted, inserted and/or added in the amino acid sequence of SEQ ID NO: 2, and having the luciferase activity; (c') a protein consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 8, and having the luciferase activity; and, (d') a protein consisting of an amino acid sequence

encoded by a polynucleotide that hybridizes under stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 7, and having the luciferase activity; and the like.

The term "hybridizes under stringent conditions" will be 5 described later.

As used herein, the range of "at least one" in the "amino acid sequence in which at least one amino acid residue is deleted, substituted, inserted and/or added" includes, for example, 1 to 17, 1 to 10, 1 to 9, 1 to 8, 1 to 7, 1 to 6 (1 to 10 several), 1 to 5, 1 to 4, 1 to 3, 1 or 2, and 1. A smaller number of the amino acids deleted, substituted, inserted and/or added is more preferable in general. Two or more types of modifications from the above deletions, substitutions, insertions and additions in amino acid residues may occur concurrently. 15 Such proteins may be obtained by using the site-specific mutagenesis described in, for example, MOLECULAR CLONING, 3rd. edition; Current Protocols in Molecular Biology; Nuc. Acids. Res., 10, 6487 (1982); Proc. Natl. Acad. Sci. USA, 79, 6409 (1982); Gene 34, 315 (1985); Nuc. Acids. 20 Res., 13, 4431 (1985); or Proc. Natl. Acad. Sci. USA, 82, 488 (1985), etc.

(2) Luciferin

The luciferin used in the present invention refers to a substance which is oxidized to oxyluciferin in the presence of 25 ods, insulin measurement kits commercially available, etc. oxygen by the catalytic action of a luciferase. This oxidization reaction by the oxygen of luciferin which is catalyzed by a luciferase is called a luciferin-luciferase reaction. The oxyluciferin formed by the luciferin-luciferase reaction is formed in an excited state, and emission of light occurs during a 30 transition to the ground state. That is, luciferin is a luminescent substrate for the luciferin-luciferase reaction.

The luciferin used in the present invention has a substrate specificity to the luciferin-luciferase reaction and is appropriately chosen depending on the luciferase used. For example, 35 where Gaussia luciferase used as the luciferin, coelenterazine (CTZ) is employed. Coelenterazine is used also where other coelenterazine-type luciferases (e.g., Renilla luciferase, Pleuromamma luciferase, Metridia luciferase, Oplophorus luciferase, etc.).

(3) Preproinsulin, Proinsulin and Insulin

Insulin is a peptide hormone composed of an A chain consisting of 21 amino acid residues and a B chain consisting of 30 amino acid residues, and plays an important role in the regulation of glucose metabolism. Insulin is formed by enzy- 45 matic cleavage of proinsulin as a precursor composed of 86 amino acid residues at the two basic dipeptide portions to produce its A and B chains and then a disulfide bond formation between the two portions. Proinsulin is produced as preproinsulin which is composed of a prepeptide (signal pep- 50 tide) with 24 amino-acid residues at the N terminus but, immediately after translocation through the rough endoplasmic reticulum, the prepeptide is cleaved to produce proinsu-

protein consisting of the amino acid sequence of SEQ ID NO: 10 and a protein having substantially the same activity or function as the protein consisting of the amino acid sequence of SEQ ID NO: 10.

The term "substantially the same activity or function" is 60 used to mean, for example: (i) a function that the substance formed from the protein above by cleavage of a signal peptide, cleavage at the two basic dipeptide portions and disulfide bond formation at the two portions lowers blood glucose levels; (ii) the activity of lowering blood glucose levels by the substance formed from the protein above through cleavage of a signal peptide, cleavage at the two basic dipeptide portions

18

and disulfide bond formation at the two portions is 1/4 or more, preferably 1/3 or more, more preferably 1/2 and most preferably 1/1.5 or more, than the activity of lowering blood glucose levels by insulin formed from the protein consisting of the amino acid sequence of SEQ ID NO: 10, or the like. The substantially the same activity or function is sometimes also referred to as the "insulin activity." The blood glucose-lowering activity described above can be assayed, for example, by administering a test compound to a test animal (mice, rats, spontaneously diabetic model KKAy mice, etc.) and measuring blood glucose levels before and after administration using known blood glucose assay methods, blood glucose meters and blood glucose kits commercially available, etc. Test animals which can be used are, for example, KKAy/Ta mice (CLEA Japan, Inc.). Blood glucose meters, blood glucose kits, etc. which can be used include, for example, Glucocard Diameter α , Diasensor (Arkray Co.), blood glucose meter Glutest Ace (GT-1640) (Sanwa Kagaku Kenkyusho Co.), blood glucose meter Dexter Z (Bayer Medical Ltd.), blood glucose meter Antosense II (Bayer-Sankyo), blood glucose meter Acucheck Comfort (Roche Diagnostics), Glucose CII-Test Wako (Wako Pure Chemicals), and the like.

The level of insulin can be measured using known meth-For example, the level of insulin can be measured using an insulin measurement kit commercially available (Morinaga Institute of Biological Science, Inc.), etc.

More specifically, the preproinsulin used in the present invention includes, for example, (a) a protein comprising the amino acid sequence of SEQ ID NO: 10; (b) a protein consisting of an amino acid sequence in which at least one amino acid residue is deleted, substituted, inserted and/or added in the amino acid sequence of SEQ ID NO: 10, and having the insulin activity; (c) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 10, and having the insulin activity; (d) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by 40 a polynucleotide that hybridizes under stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 9, and having the insulin activity, etc.

Preferably, the preproinsulin includes the signal peptide of preproinsulin and one of the polypeptides described in (i) through (1).

(i) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 4;

(j) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 4, and having the

(k) a polypeptide comprising a polypeptide consisting of The preproinsulin used in the present invention includes a 55 an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 4, and having the insulin

> (1) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 3, and having the insulin activity.

> Preferably, the signal peptide of preproinsulin is a peptide consisting of the amino acid sequence of SEQ ID NO: 6.

> The proinsulin used in the present invention includes a protein consisting of the amino acid sequence of SEQ ID NO:

10 and a protein having substantially the same activity or function as the protein consisting of the amino acid sequence of SEQ ID NO: 10.

The term "substantially the same activity or function" is used to mean, for example, (i) a function that the substance formed from the protein above by cleavage of a signal peptide, cleavage at the two basic dipeptide portions and disulfide bond formation at the two portions lowers blood glucose levels; (ii) the activity of lowering blood glucose levels by the substance formed from the protein above through cleavage of a signal peptide, cleavage at the two basic dipeptide portions and disulfide bond formation at the two portions is ½4 or more, preferably ½ or more, more preferably ½ and most preferably 1/1.5 or more, than the activity of lowering blood glucose levels by insulin formed from the protein consisting of the amino acid sequence of SEQ ID NO: 10, or the like.

(4) Fusion Protein of the Preproinsulin and the Luciferase

As used herein, the fusion protein of preproinsulin and the luciferase refers to a fusion protein of the preproinsulin 20 described above and the luciferase described above.

As described above, where the luciferase is a secretory luciferase, the luciferase may have a signal peptide or may lack a signal peptide.

In the specification, the fusion protein of preproinsulin and 25 the luciferase is sometimes also referred to as the "fusion protein of the present invention."

The fusion protein of the present invention comprises any one of the polypeptides described in any one of (m) through (p) below:

(m) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 12;

(n) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues 35 in the amino acid sequence of SEQ ID NO: 12, and having the insulin activity;

(o) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 12, and having the 40 insulin activity; and,

(p) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to 45 the nucleotide sequence of SEQ ID NO: 11, and having the insulin activity.

(5) Polynucleotide Encoding the Fusion Protein of the Present Invention

In the specification, the polynucleotide encoding the fusion 50 protein of the preproinsulin and the luciferase described above is sometimes also referred to as the "polynucleotide encoding the fusion protein of the present invention."

In the specification, the polynucleotide encoding the fusion protein of the present invention is sometimes also referred to 55 as the "polynucleotide of the present invention."

The polynucleotide of the present invention may be any polynucleotide as far as it is a polynucleotide comprising the nucleotide sequence encoding the fusion protein of the present invention, and is preferably a DNA.

The DNA includes genomic DNA, genomic DNA library, cDNA derived from the cells or tissues, cDNA library derived from the cells and tissues described above, synthetic DNA, etc. The vector used for the library may be any of bacteriophage, plasmid, cosmid and phagemide, and is not particularly limited. The DNA can also be directly amplified by reverse transcriptase polymerase chain reaction (hereinafter abbrevi-

20

ated as RT-PCR) employing a total RNA or a mRNA fraction prepared from the cells or tissues described above.

The polynucleotide of the invention includes: (m) a polynucleotide encoding a polypeptide consisting of the amino acid sequence of SEQ ID NO: 12; (n) a polynucleotide encoding a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 12, and having the insulin activity; (o) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 12, and having the insulin activity; and, (p) a polynucleotide encoding a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 11, and having the insulin activity. The polynucleotide of the present invention is preferably (p2) a polynucleotide further comprising the nucleotide sequence of SEQ ID NO: 11, and more preferably (p3) a polynucleotide consisting of the nucleotide sequence of SEQ ID NO: 11.

As used herein, the "polynucleotide (e.g., a DNA) that hybridizes under stringent conditions" means a polynucleotide (e.g., a DNA) which is obtained by colony hybridization, plaque hybridization, Southern blot hybridization or the like using as a probe the whole or part of the target polynucleotide (for example, a polynucleotide (e.g., a DNA) consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 11 (for example, a polynucleotide encoding the amino acid sequence encoding the amino acid sequence of SEQ ID NO: 12 (e.g., a DNA)). Specifically, the polynucleotide includes a polynucleotide which can be identified by performing hybridization at 65° C. in the presence of 0.7 to 1.0 mol/L NaCl using a filter on which the DNA derived from colony or plaque is immobilized, and then washing the filter in 0.1 to 2×SSC (saline-sodium citrate) solution (1×SSC solution is composed of 150 mmol/L sodium chloride and 15 mmol/L sodium citrate) at 65° C.

Hybridization may be carried out based on the methods described in laboratory manuals such as Sambrook J. et al., Molecular Cloning: A Laboratory Manual, Third Edition, Cold Spring Harbor Laboratory Press (2001) (hereinafter briefly referred to as Molecular Cloning, 3rd edition), Ausbel F. M. et al., Current Protocols in Molecular Biology, Supplement 1-38, John Wiley and Sons (1987-1997), Glover D. M. and Hames B. D., DNA Cloning 1: Core Techniques, A Practical Approach, Second Edition, Oxford University Press (1995), etc.

As used herein, the "stringent conditions" may be any of low-stringent conditions, medium-stringent conditions and high-stringent conditions. The "low-stringent conditions" are, for example, 5×SSC, 5×Denhardt's solution, 0.5% (w/v) SDS, 50% (v/v) formamide and 32° C. The "medium-stringent conditions" are, for example, 5×SSC, 5×Denhardt's solution, 0.5% (w/v) SDS, 50% (v/v) formamide and 42° C. The "high-stringent conditions" are, for example, 5×SSC, 5×Denhardt's solution, 0.5% (w/v) SDS, 50% (v/v) formamide and 50° C. As the conditions become severer, complementation required for duplex formation becomes higher. Specifically, under these conditions, for example, as the temperature is higher, a DNA with higher homology is expected to be obtained efficiently, although multiple factors are involved in the hybridization stringency including temperature, probe concentration, probe length, ionic strength, time,

salt concentration and the like. Those skilled in the art may achieve similar stringency by appropriately choosing these factors

When a commercially available kit is used for hybridization, for example, Alkphos Direct Labeling Reagents (manufactured by Amersham Pharmacia) can be used. In this case, according to the attached protocol, a membrane is incubated with a labeled probe overnight, the membrane is washed with a primary wash buffer containing 0.1% (w/v) SDS at 55° C. and then the hybridized DNA can be detected.

Other polynucleotides that can be hybridized include DNAs having an identity of approximately 80% or higher, 85% or higher, 88% or higher, 90% or higher, 92% or higher, 95% or higher, 97% or higher. 98% or higher, 99% or higher, 99.3% or higher, 99.5% or higher, 99.7% or higher, 99.8% or higher or 99.9% or higher, to the nucleotide sequence of the target polynucleotide (e.g., DNA), as calculated by homology search software, such as FASTA and BLAST using default parameters.

The polynucleotide encoding a protein consisting of an amino acid sequence in which at least one amino acid residue is deleted, substituted, inserted and/or added in a given amino acid sequence may be obtained, using a site-specific mutagenesis technique (cf., for example, Gotoh, T. et al., Gene, 152, 271-275 (1995); Zoller, M. J. and Smith, M., 25 Methods Enzymol., 100, 468-500 (1983); Kramer, W. et al., Nucleic Acids Res., 12, 9441-9456 (1984); Kramer, W. and Fritz, H. J., Methods Enzymol., 154, 350-367 (1987); Kunkel, T. A., Proc. Natl. Acad. Sci. USA, 82, 488-492 (1985); Kunkel, Methods Enzymol., 85, 2763-2766 (1988), 30 etc.) and methods using amber mutation (cf., for example, the gapped duplex method in Nucleic Acids Res., 12, 9441-9456 (1984), etc.).

Alternatively, a mutation may also be introduced onto the polynucleotide by PCR using a set of primers bearing on each 35 5' end a sequence in which the target mutation (deletion, addition, substitution and/or insertion) has been introduced (cf., for example, Ho, S. N. et al., Gene, 77, 51 (1989), etc.).

Specific examples of the polynucleotide (i.e., polynucleotide encoding the fusion protein of the preproinsulin and the 40 luciferase) of the present invention include:

- (i) a polynucleotide encoding the fusion protein of preproinsulin and a secretory luciferase;
- (ii) a polynucleotide encoding the fusion protein of preproinsulin and *Gaussia* luciferase;
- (iii) a polynucleotide encoding the fusion protein of preproinsulin and the protein (*Gaussia* luciferase) described in any one of (a) through (d) below;
- (a) a protein comprising the amino acid sequence of SEQ ID NO: 8;
- (b) a protein consisting of an amino acid sequence in which at least one amino acid residue is deleted, substituted, inserted and/or added in the amino acid sequence of SEQ ID NO: 8, and having the luciferase activity;
- (c) a protein consisting of an amino acid sequence having at 55 least 90% identity to the amino acid sequence of SEQ ID NO: 8, and having the luciferase activity; and,
- (d) a protein consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under stringent conditions to a polynucleotide consisting of a nucleotide sequence 60 complementary to the nucleotide sequence of SEQ ID NO: 7, and having the luciferase activity;
- (iv) a polynucleotide encoding the fusion protein of preproinsulin and the protein (*Gaussia* luciferase) described in any one of (a) through (d) below:
- (a) a protein comprising the amino acid sequence of SEQ ID NO: 8;

22

- (b) a protein consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 8, and having the luciferase activity;
- (c) a protein consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 8, and having the luciferase activity;
- (d) a protein consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 7, and having the luciferase activity;
- (v) a polynucleotide encoding the fusion protein of the polypeptide (preproinsulin) described in any one of (e) through (h) below and the luciferase described in any one of (i) through (iv) above:
- (e) a polypeptide consisting of the amino acid sequence of SEO ID NO: 10:
- (f) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 10, and having the insulin activity;
- (g) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 10, and having the insulin activity; and,
- (h) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 9, and having the insulin activity;
- (vi) a polynucleotide encoding the fusion protein between a polypeptide (preproinsulin), which consists of the signal peptide of preproinsulin and the polypeptide described in any one of (i) through (l) below, and the luciferase described in any one of (i) through (iv) above:
- (i) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 4;
- (j) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 4, and having the insulin activity:
- (k) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the 50 amino acid sequence of SEQ ID NO: 4, and having the insulin activity; and,
 - (l) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 3, and having the insulin activity.
 - (vii) a polynucleotide encoding the fusion protein between a polypeptide (preproinsulin), which consists of a polypeptide (signal peptide of preproinsulin) consisting of the amino acid sequence of SEQ ID NO: 6 and the polypeptide described in any one of (i) through (l) below, and the luciferase described in any one of (1) through (4) above:
 - (i) a polypeptide consisting of the amino acid sequence of SEO ID NO: 4;
 - (j) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, inser-

tion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 4, and having the insulin activity;

(k) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the 5 amino acid sequence of SEQ ID NO: 4, and having the insulin activity; and,

(l) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucle- 10 otide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 3, and having the insulin activity;

(viii) a polynucleotide described in any one of (i) through (vii) above, encoding the polypeptide described in any one of (m) 15 through (p) below:

(m) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 12;

(n) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, 20 insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 12, and having the insulin activity;

(o) a polypeptide comprising a polypeptide consisting of amino acid sequence of SEQ ID NO: 12, and having the insulin activity; and,

(p) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 11, and having the insulin activity; and the like.

The present invention can further provide the recombinant vector and transformant comprising the polynucleotide 35 described above.

(6) Recombinant Vector

The vector capable of expressing the fusion protein of the present invention can be obtained by ligating (inserting) the polynucleotide encoding the fusion protein of the preproin- 40 sulin and the luciferase (the polynucleotide of the invention (DNA)) to a suitable vector.

As used herein, the recombinant vector bearing the polynucleotide of the invention is sometimes also referred to as the "recombinant vector of the invention."

More specifically, the recombinant vector may be obtained by cleaving a purified polynucleotide (DNA) with a suitable restriction enzyme, then inserting the resulting fragment into a restriction enzyme site or multicloning site on a suitable vector, and ligating to the vector. The vector for inserting the 50 polynucleotide of the invention includes, for example, but not limited to, plasmids, bacteriophages, animal viruses, and the like. Examples plasmids include plasmids derived from Escherichia coli (e.g., pBR322, pBR325, pUC118, pUC119, etc.), plasmids derived from Bacillus subtilis (e.g., pUB110 55 and pTP5), and plasmids derived from yeast (e.g., YEp13, YEp24 and YCp50). An example of bacteriophage is λ phage. Examples of animal viruses include retroviruses, vaccinia viruses, insect viruses (e.g., baculoviruses), etc.

The polynucleotide of the invention is generally ligated 60 downstream of the promoter in a suitable vector in an expressible form. Where the host used for transformation is an animal cell, preferred promoters are promoters from SV40, retrovirus promoters, metallothionein promoters, heat shock promoters, cytomegalovirus promoters, SRα promoters and the 65 like. Where the host is bacteria belonging to the genus Escherichia, preferred promoters include Trp promoter, T7

24

promoter, lac promoter, recA promoter, \(\lambda PL \) promoter, lpp promoter, etc. Where the host is bacteria belong to genus Bacillus, preferred promoters include SPO1 promoter, SPO2 promoter, penP promoter, etc. Where the host is a yeast, preferred promoters include PHO5 promoter, PGK promoter, GAP promoter, ADH1 promoter, GAL promoter, etc. Where the host is an insect cell, preferred promoters include polyhedrin promoter, P10 promoter, etc.

In addition to the foregoing, the recombinant vector of the invention which can be used may contain, if desired, an enhancer, a splicing signal, a poly(A) addition signal, a ribosome binding sequence (SD sequence), a selective marker and the like. Examples of selective markers include dihydrofolate reductase gene, ampicillin resistance gene, neomycin resistance gene, etc.

(7) Transformant

The transformant can be produced by introducing the recombinant vector comprising the polynucleotide of the invention (i.e., the polynucleotide encoding the fusion protein of the invention) into a suitable host.

In the specification, the transformant comprising the recombinant vector of the present invention is sometimes also referred to as the "transformant of the invention."

The host used to produce the transformant of the invention an amino acid sequence having at least 90% identity to the 25 is not particularly limited, so long as the host is capable of expressing the polynucleotide (DNA) of the invention. Examples include bacteria of the genera Escherichia, Bacillus, Pseudomonas and Rhizobium, yeasts, animal cells and insect cells. Bacteria of the genus Escherichia include, for example, Escherichia coli. Bacteria of the genus Bacillus include, for example, *Bacillus subtilis*. Bacteria of the genus Pseudomonas include, for example, Pseudomonas putida. Bacteria of the genus Rhizobium include Rhizobium meliloti. Yeasts include, for example, Saccharomyces cerevisiae and Schizosaccharomyces pombe. Animal cells include, for example, COS cells and CHO cells. Insect cells include, for example, Sf9 and Sf21.

> Among others, animal cells and cell lines derived from animal cells are preferred as the host. More preferably, the host is animal cells capable of secreting insulin, pancreatic Langerhans islet β cells and cell lines derived from these

Introduction of the recombinant vector into the host and transformation thereby may be performed by various methods generally used. These methods for introducing the recombinant vector into the host cell include the calcium phosphate method (Virology, 52, 456-457 (1973)), the lipofection method (Proc. Natl. Acad. Sci. USA, 84, 7413 (1987)), the electroporation method (EMBO J., 1, 841-845 (1982)), etc. Methods for transforming bacteria of the genus Escherichia include, for example, the methods described in Proc. Natl. Acad. Sci. USA, 69, 2110 (1972), Gene, 17, 107 (1982), etc. Methods for transforming bacteria of the genus Bacillus include, for example, the methods described in Molecular & General Genetics, 168, 111 (1979). Methods for transforming yeasts include, for example, the methods described in Proc. Natl. Acad. Sci. USA, 75, 1929 (1978). Methods for transforming animal cells include, for example, the methods described in Virology, 52, 456 (1973). Methods for transforming insect cells include, for example, the methods described in Bio/Technology, 6, 47-55 (1988). The transformant comprising the polynucleotide encoding the protein of the invention (polynucleotide of the invention) can be produced in such a way.

(8) Culture of the Transformant of the Invention

The transformant of the invention may be cultured in a conventional manner used for culturing hosts. The transfor-

mant is so cultured to produce the fusion protein of the invention, and the fusion protein of the invention is secreted into the culture broth.

The medium for culturing the transformant using bacteria of the genus Escherichia or the genus Bacillus as a host may 5 be any of a natural medium and a synthetic medium, as far as it is a medium which contains carbon sources, nitrogen sources, inorganic salts, etc. necessary for growth of the transformant, and in which the transformant can be efficiently grown. Examples of carbon sources which may be used are 10 carbohydrates such as glucose, fructose, sucrose, starch, etc.; organic acids such as acetic acid, propionic acid, etc.; alcohols such as ethanol and propanol, and the like. Examples of nitrogen sources which may be used include ammonia, ammonium salts of inorganic or organic acids such as ammonium chloride, ammonium sulfate, ammonium acetate, ammonium phosphate, etc., and other nitrogen-containing compounds, and further include peptone, meat extracts, corn steep liquor, and the like. Examples of inorganic salts include monobasic potassium phosphate, dibasic potassium phos- 20 phate, magnesium phosphate, magnesium sulfate, sodium chloride, ferrous sulfate, manganese sulfate, copper sulfate, calcium carbonate, etc. Antibiotics such as ampicillin or tetracycline may be added to the medium during culture, depending on necessity. Where the transformant transformed 25 by an expression vector using an inducible promoter as the promoter, if necessary, an inducer may also be added to the medium. For example, isopropyl-β-D-thiogalactopyranoside (IPTG), etc. may be added to the medium when a transformant transformed by an expression vector using a Lac pro- 30 moter is cultured, and indoleacrylic acid (IAA), etc. may be added to the medium when a transformant transformed by an expression vector using a trp promoter is cultured.

When the host is bacteria of the genus *Escherichia*, incubation is performed generally at approximately 15 to 43° C. 35 for approximately 3 to 24 hours. If necessary, aeration and agitation may be applied. When the host is bacteria of the genus *Bacillus*, incubation is performed generally at approximately 30 to 40° C. for approximately 6 to 24 hours. If necessary, aeration and agitation may be applied.

Media for culturing the transformant when the host is yeast include Burkholder's minimal medium (Proc. Natl. Acad. Sci. USA, 77, 4505 (1980)) and an SD medium containing 0.5% (w/v) casamino acids (Proc. Natl. Acad. Sci. USA, 81, 5330 (1984)). Preferably, the pH of the medium is adjusted to 45 approximately 5 to 8. Culture is performed generally at approximately 20 to 35° C. for approximately 24 to 72 hours. If necessary, aeration and agitation may be applied.

Media for culturing the transformant when the host is an animal cell include MEM medium containing approximately 50 5 to 20% (v/v) fetal calf serum (Science, 122, 501 (1952)), DMEM medium (Virology, 8, 396 (1959)), etc. Preferably, the pH of the medium is adjusted to approximately 6 to 8. Culture is performed generally at approximately 30 to 40° C. for approximately 15 to 60 hours. If necessary, aeration and 55 agitation may be applied.

Media for culturing the transformant when the host is an insect cell include Grace's insect medium (Nature, 195, 788 (1962)) to which additives such as 10% (v/v) immobilized bovine serum are suitably added. Preferably, the pH of the 60 medium is adjusted to approximately 6.2 to 6.4. Culture is performed generally at approximately 27° C. for approximately 3 to 5 hours. If necessary, aeration and agitation may be applied.

The transformant of the invention is cultured under the 65 culture conditions described above (i) in the absence of a test compound or (ii) in the presence of a test compound.

26

Examples of the test compound are peptides, proteins, nonpeptide compounds, synthetic compounds, fermentation products, cell extracts, plant extracts, animal tissue extracts, blood plasma and so on. The test compound may form a salt, and a pharmacologically acceptable salt is preferred as the salt of the test compound. Examples of pharmacologically acceptable salts include metal salts, ammonium salts, salts with organic bases, salts with inorganic acids, salts with organic acids, salts with basic or acidic amino acids, and the like. Examples of metal salts include alkali metal salts such as sodium salts, potassium salts, etc.; alkaline earth metal salts such as calcium salts, magnesium salts, barium salts, etc.; aluminum salts, and the like. Examples of salts with organic bases include salts with trimethylamine, triethylamine, pyridine, diethanolamine, triethanolamine, cyclohexylamine, dicyclohexylamine, N,N-dibenzylethylenediamine, Examples of salts with inorganic acids include salts with hydrochloric acid, hydrobromic acid, nitric acid, sulfuric acid, phosphoric acid, etc. Examples of salts with organic acids include salts with formic acid, acetic acid, trifluoroacetic acid, propionic acid, phthalic acid, fumaric acid, oxalic acid, tartaric acid, maleic acid, citric acid, succinic acid, malic acid, methanesulfonic acid, benzoic acid, benzenesulfonic acid, p-toluenesulfonic acid, and the like. Examples of salts with basic amino acids include salts with arginine, lysine, ornithine, etc. Examples of salts with acidic amino acids include salts with aspartic acid, glutamic acid, and the like. Step (b)

(8) Expression (Formation) of the Fusion Protein of the Invention and its Extracellular Secretion

As described above, the transformant of the invention is cultured, whereby the fusion protein of the invention can be expressed (produced) in the transformant and secreted outside the cells (into a medium).

Luciferin which can cause the luciferin-luciferase reaction specifically with the luciferase in the fusion protein of the present invention is previously added to a medium. When the fusion protein of the present invention is secreted extracellularly, the luciferin-luciferase reaction with the luciferin in the medium is caused to emit light. This luminescence is observed, detected or measured, and the localization (distribution), secretion, quantification, etc. of the fusion protein of the present invention can be observed, detected or measured.

45 Step (c)

(9) Luminescence Imaging

As used herein, luminescence imaging means the observation, detection or measurement of luminescence.

The transformant of the present invention is capable of expressing and secreting the fusion protein of the present invention. The fusion protein of the present invention can cause the luciferin-luciferase reaction between the luciferase in the fusion protein and the luciferin specific to the luciferase above. As described above, light emits by the luciferin-luciferase reaction. By observing, detecting or measuring this luminescence, the localization (distribution), secretion, quantification, etc. of the fusion protein of the present invention can be observed, detected or measured.

Specifically, the luminescence imaging can be performed using an optical detection device (e.g., a camera), including a camera such as a CCD camera, a photon counting camera, etc. (an electron camera), which detects light electronically, a camera such as a film camera, etc. (a film camera or a silver halide camera), which detects light scientifically. A monochrome CCD camera, a 3CCD camera, a single CCD color camera, a digital CCD camera, etc. can be used. Furthermore, a ultra-weak-light detectable CCD camera may also be used

and includes an electronically cooled CCD, a liquid nitrogencooled CCD, a CCD with imaging intensifier, etc.

Preferably, the ultra-weak-light detectable CCD imaging camera is an EM-CCD camera (e.g., model C9100-13 manufactured by Hamamatsu Photonics K. K., model C9100-14⁵ model manufactured by Hamamatsu Photonics K. K., model OuantEM manufactured by Princeton Instrument Inc., Evolve manufactured by Photometrics Corp., model Night-OWL II LB983 em100 manufactured by Berthold Inc., etc.). The photon counting camera model which may be used includes a device commercially available such as a VIM camera manufactured by Hamamatsu Photonics K. K. The VIM camera manufactured by Hamamatsu Photonics K. K. is equipped with an optical amplifier and a CCD camera.

The luminescence imaging may be performed using a microscope in addition to the optical detection device, preferably using a microscope. Microscopes commercially available may also be used and include, e.g., an inverted microscope such as models IX71 and IX81 manufactured by 20 Step (a') Olympus Corporation.

The luminescence imaging may also be performed further using an image intensifier (optical amplifier device) such as an optical amplifier tube, a semiconductor optical amplifier device, etc., in addition to the optical detection device, pref-25 erably using an optical amplifier device. The optical amplifier device which may be used includes a semiconductor photodetector (e.g., a GaAsP (gallium arsenic phosphide) image intensifier, etc.), a photomultiplier, etc. The optical amplifier device is preferably a GaAsP image intensifier (e.g., model C8600-04 manufactured by Hamamatsu Photonics K. K.). Step (d)

(10) Screening of the Compound that Promotes (or Regulates) the Secretion of the Fusion Protein of the Invention or 35 Insulin Secretion from Cells

As described above, the transformant of the present invention is cultured in the absence of a test compound and in the presence of the test compound and levels of secretion or the like of the fusion protein of the invention are measured in the 40 respective cases by observing the luminescence caused by the luciferin-luciferase reaction. By comparing the levels of secretion or the like of the fusion protein of the invention in the absence and the presence of the test compound, a substance that promotes (or regulates) the secretion or the like of 45 the fusion protein of the invention or insulin from cells can be screened.

More specifically, the substance that promotes (or regulates) the secretion of the fusion protein of the invention or insulin from cells can be screened by measuring the levels of secretion, etc. of the fusion protein of the present invention when the transformant of the present invention is cultured (i) in the absence of a test compound and the levels of secretion, etc. of the fusion protein of the present invention when the transformant of the present invention is cultured (ii) in the presence of the test compound, and comparing the secretion levels by luminescence imaging.

More specifically, for example, in the case of (ii) above, a test compound which promotes the level of secretion of the fusion protein of the invention by about 10% or more, preferably about 20% or more, more preferably about 30% or more, much more preferably about 40% or more and most preferably about 50% or more, when compared to the case of (i) above, can be selected as a substance that promotes (or 65 regulates) the secretion of the fusion protein or the invention or the insulin secretion, etc.

28

Screening Method 2:

The method of screening a substance that regulates the secretion of MMP-2 from cells, which comprises Steps (a') through (d') below:

Step (a'): the step of culturing a cell transformed by the polynucleotide encoding the fusion protein of the luciferase and pro-MMP-2 in a medium in the presence or absence of a test compound;

Step (b'): the step of expressing the fusion protein of the present invention to secrete the fusion protein of the invention into a luciferin-containing medium (outside the cells).

Step (c'): the step of observing the fusion protein secreted extracellularly by luminescence imaging; and,

Step (d'): the step of comparing the expression and/or activity of the fusion protein or the cancer metastasis suppressing activity, etc. in the presence of or in the absence of a test compound.

The steps are explained below in detail.

(1') Luciferase

The luciferase used in the present invention is as described in the screening method 1 above. The luciferase used includes those listed in the screening method 1 above and preferred examples are the same as well.

(2') Luciferin

The luciferin used in the present invention is as described in the screening method 1 above. The luciferin used includes those listed in the screening method 1 above and preferred examples are the same as well.

(3') pro-MMP-2 and MMP-2

Matrix metalloproteinase (MMP)-2 is a zinc-dependent endopeptidase involved in the degradation and repair of the major components of extracellular matrix and connective tissues. MMP-2 has an activity of degrading type IV collagen as the main components of the basal membrane and denatured collagen (gelatin). MMP-2 also has an activity of disrupting vascular membranes mainly composed of type IV collagen. Based on these activities, MMP-2 plays an important role in tumor metastasis.

MMP-2 is synthesized as pro-MMP-2 in which the signal peptide is appended to the N terminus. pro-MMP-2 is activated through cleavage of the signal peptide to form MMP-2.

The pro-MMP-2 used in the present invention comprises a protein consisting of the amino acid sequence of SEQ ID NO: 18 and a protein having substantially the same activity or function as the protein consisting of the amino acid sequence of SEQ ID NO: 18.

The term "substantially the same activity or function" is used to mean that, for example: (i) the activity of degrading the active protein type IV collagen produced from the protein above via cleavage of the signal peptide or the activity of degrading denatured collagen (gelatin) is 1/4 or more, preferably $\frac{1}{3}$ or more, more preferably $\frac{1}{2}$ and most preferably $\frac{1}{1.5}$ or more, than the type IV collagen degradation activity or denatured collagen (gelatin) degradation activity of MMP-2 produced from the protein consisting of the amino acid sequence of SEQ ID NO: 8, or the like. The substantially the same activity or function is sometimes referred to as "a MMP-2 activity." The type IV collagen degradation activity or denatured collagen (gelatin) degradation activity described above can be assayed by adding the protein described above or MMP-2 to type IV collagen or denatured collagen (gelatin) to perform their degradation reaction for a given period of time and quantitatively determining the type IV collagen or denatured collagen (gelatin) remained using HPLC, gel electrophoresis, etc.

More specifically, the pro-MMP-2 used in the present invention includes, for example, (a') a protein comprising the amino acid sequence of SEQ ID NO: 18; (b') a protein consisting of an amino acid sequence in which at least one amino acid residue is deleted, substituted, inserted and/or added in the amino acid sequence of SEQ ID NO: 18, and having the MMP-2 activity; (c') a protein consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 18, and having the MMP-2 activity; and, (d') a protein consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 17, and having the MMP-2 activity, and the like.

The pro-MMP-2 used in the present invention further includes, for example, (a') a protein comprising the amino acid sequence of SEQ ID NO: 18; (b') a protein consisting of an amino acid sequence in which at least one amino acid residue is deleted, substituted, inserted and/or added in the 20 amino acid sequence of SEQ ID NO: 18 and having the MMP-2 activity in the form of the active protein which is obtained by cleavage of the signal peptide from the amino acid sequence above; (c') a protein consisting of an amino acid sequence having at least 90% identity to the amino acid 25 sequence of SEQ ID NO: 18, and having the MMP-2 activity in the form of the active protein which is obtained by cleavage of the signal peptide from the amino acid sequence above; and, (d') a protein consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under stringent 30 conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 17, and having the MMP-2 activity in the form of the active protein which is obtained by cleavage of the signal peptide from the amino acid sequence above, and the like.

Preferably, the pro-MMP-2 consists of the signal peptide of pro-MMP-2 and the protein described in any one of (i') through (P) below:

- (i') a protein consisting of the amino acid sequence of SEQ ID NO: 14;
 - (j') a protein comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 14, and having the MMP-2 activity;
- (k') a protein comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 4, and having the MMP-2 activity; and,
 - (I') a protein comprising a polypeptide consisting of an 50 amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 3, and having the MMP-2 activity.

Preferably, the signal peptide of pro-MMP-2 is a peptide consisting of the amino acid sequence of SEQ ID NO: 16. (4') Fusion Protein Between pro-MMP-2 and Luciferase

As used herein, the fusion protein of pro-MMP-2 and the luciferase refers to the pro-MMP-2 described above and the 60 luciferase described above.

As described above, where the luciferase is a secretory luciferase, the luciferase may have a signal peptide or may lack a signal peptide.

In the specification, the fusion protein of pro-MMP-2 and 65 the luciferase is sometimes also referred to as the "fusion protein of the present invention."

The fusion protein of the present invention comprises the protein described in any one of (m') through (p') below:

- (m') a protein consisting of the amino acid sequence of SEQ ID NO: 20;
- (n') a protein comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 20, and having the MMP-2 activity;
- (o') a protein comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 20, and having the MMP-2 activity; and,
- (p') a protein comprising a polypeptide consisting of an 15 amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 19, and having the MMP-2 activity.
 - (5') Polynucleotide Encoding the Fusion Protein of pro-MMP-2 and the Luciferase

In the specification, the polynucleotide encoding the fusion protein of pro-MMP-2 and the luciferase is sometimes also referred to as the polynucleotide encoding the fusion protein of the present invention.

In the specification, the polynucleotide encoding the fusion protein described above is sometimes also referred to as the "polynucleotide of the present invention."

The polynucleotide of the present invention may be any polynucleotide as far as it is a polynucleotide comprising the nucleotide sequence encoding the fusion protein of the present invention, and is preferably a DNA.

The polynucleotide of the present invention includes (m') a polynucleotide encoding the protein consisting of the amino 35 acid sequence of SEQ ID NO: 20; (n') a polynucleotide encoding the protein consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of one to several amino acid residues in the amino acid sequence of SEQ ID NO: 20, and having the MMP-2 activity; (o') a protein comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 20, and having the MMP-2 activity; and, (p') a polynucleotide encoding a protein comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEO ID NO: 19, and having the MMP-2 activity. The polynucleotide of the present invention is preferably (p2') a polynucleotide comprising the nucleotide sequence of SEQ ID NO: 19, and more preferably (p3') a polynucleotide consisting of the nucleotide sequence of SEQ ID NO: 19.

According to the present invention, the recombinant vector and transformant bearing the polynucleotide of the present invention described above can be further provided.

(6') Recombinant Vector

A vector capable of expressing the fusion protein of the invention can be acquired by ligating (inserting) the polynucleotide encoding the fusion protein of pro-MMP-2 and the luciferase (polynucleotide of the present invention (DNA)) to or into a suitable vector.

In the specification, the recombinant vector bearing the polynucleotide of the present invention is sometimes also referred to as the "recombinant vector of the present invention."

More specifically, the recombinant vector of the present invention can be produced as described above.

(7') Transformant

The transformant can be prepared by introducing into a suitable host the recombinant vector bearing the polynucleotide encoding the fusion protein of this pro-MMP-2 and the luciferase (namely, the polynucleotide encoding the fusion protein of the present invention) thus produced.

In the specification, the transformant bearing the recombinant vector of the present invention is sometimes also referred to as the "transformant of the present invention."

More specifically, the transformant of the present invention can be prepared as described above.

(8') Culture of the Transformant of the Invention

The transformant of the present invention can be cultured in a conventional manner used in the culture of a host, as described above. By the culture, the fusion protein of the invention is produced and the fusion protein of the present invention is secreted into the culture broth.

Step (c')

(9') Luminescence Imaging

The luminescence imaging can be performed as described above.

Step (d')

(10) Screening of a Compound that Promotes (or Regulates) the Secretion of the Fusion Protein of the Invention or MMP-2 Secretion from Cells

As described above, the transformant of the present invention is cultured in the absence of a test compound and in the presence of the test compound and levels of secretion or the like of the fusion protein of the invention are measured in the respective cases by observing the luminescence caused by the luciferin-luciferase reaction. By comparing the levels of secretion or the like of the fusion protein of the invention in the absence of the test compound and the presence of the test compound, a substance that promotes (or regulates) the secretion or the like of the fusion protein of the invention or MMP-2 from cells can be screened.

More specifically, the substance that promotes (or regulates) the secretion of the fusion protein of the invention or MMP-2 from cells can be screened by measuring the levels of secretion, etc. of the fusion protein of the present invention when the transformant of the present invention is cultured (i') uin the absence of a test compound and the levels of secretion, etc. of the fusion protein of the present invention when the transformant of the present invention is cultured (ii') in the presence of the test compound, and comparing the secretion levels by luminescence imaging.

More specifically, for example, in the case of (ii') above, a test compound which promotes the level of secretion of the fusion protein of the invention by about 10% or more, preferably about 20% or more, more preferably about 30% or more, much more preferably about 40% or more and most 50 preferably about 50% or more, when compared to the case of (i') above, can be selected as a substance that promotes (or regulates) the secretion of the fusion protein or the invention or the MMP-2 secretion, etc.

EXAMPLES

Example 1-1

Construction of Fusion Protein Between Human Preproinsulin (hINS) and *Gaussia* Luciferase (GLase)

The expression vector pcDNA3-hINS-GLuc to express the fusion protein between human preproinsulin and *Gaussia* 65 luciferase was prepared according to the following procedures.

32

The BamHI-EcoRI cDNA fragment encoding human preproinsulin was obtained by PCR using as a template IMAGE cDNA clone 3950204 (cf. http://image.llnl.gov/image/html/ vectors.shtml). Using KOD-plus-DNA polymerase (manufactured by Toyobo Co., Ltd.) and primers hINS-P1 (5'ctc GGATCC AGCCACC ATG GCC CTG TGG ATG CGC CT 3'; BamHI recognition site underlined) (SEQ ID NO: 21) and hINS-P2 (5' ctt GAATTC GTT GCA GTA GTT CTC CAG CTG 3'; EcoRI recognition site underlined) (SEQ ID NO: 22), PCR was carried out under the following cycle conditions: 25 cycles of 15 secs/96° C., 15 secs/55° C. and 45 secs/68° C. The DNA fragment obtained was digested with BamHI and EcoRI and then inserted into the BamHI/EcoRI site of pcDNA3-GLuc-BE vector (described in FEBS Letters, 581, 4551-4556 (2007)) to construct expression vector pcDNA3hINS-GLuc.

The protein expressed by the expression vector pcDNA3-hINS-GLuc obtained is the fusion protein of insulin signal peptide sequence, proinsulin and *Gaussia* luciferase.

Example 1-2

Construction of a Transformant Using Expression Vector pcDNA3-hINS-GLuc and Assessment of the Transformant

Mouse pancreatic β cells, MIN6 strains, were cultured in high glucose DMEM (manufactured by Sigma) supplemented with 10% fetal calf serum (manufactured by Invitrogen) and 50 μM of 2-mercaptoethanol.

The expression vector obtained in EXAMPLE 1-1 was transfected to the MIN6 cell line using LipofectAMINE 2000 (manufactured by Invitrogen). After the transfection, the cells were incubated in a CO₂ incubator at 37° C. for 24 hours.

The secretion expression of the fusion *Gaussia* luciferase protein in the transformant was confirmed by measuring the luminescence activity with a Luminometer AB2200 (manufactured by Atto Corporation) using coelenterazine as a luminescent substrate, in accordance with the method described in Biochem. Biophys. Res. Commun., 365, 96-101 (2007). As a result, the luminescence activity in the cultured medium by high glucose (20 mM glucose) stimulation to the transformed MIN6 cells increased by about 7 times as compared to that prior to the treatment (FIG. 1). This revealed that insulin secretion can be assessed in this expression vector pcDNA3-hINS-GLuc-animal culture cell system.

Example 1-3

Imaging of the Extracellular Secretion of Insulin by the Expression of Human Insulin-*Gaussia* Luciferase Fusion Protein

The MIN6 cells to visualize insulin secretion were cultured 55 on a 35 mm glass-bottom culture dish coated with poly-D-lysine (manufactured by Mat-Tek).

An EM-CCD camera was used to visualize insulin secreted from cells by the addition of and stimulation with high concentration glucose using a luminescence signal from the insu60 lin-GLase fusion protein.

More specifically, the MIN6 cell line transformed was rinsed 3 times with 3 ml of phosphate buffered saline (PBS) and soaked with 1 ml of a buffer containing 3 µg/ml of coelenterazine. The buffer used was low glucose-modified KRHB buffer (Krebs-Ringer Hepes buffer: 130 mM NaCl, 4.7 mM KCl, 1.2 mM KH₂PO₄, 1.2 mM MgSO₄, 1.5 mM CaCl₂, 10 mM Hepes (pH 7.4), containing 2 mM glucose).

After rinsing 1 ml of the buffer above, luminescence video images of the cells in low-glucose KRHB buffer (2 mM glucose) containing 3 µg/ml of coelenterazine were obtained. In order to acquire luminescence video images of the same cells as described above in high-glucose buffer, 1 ml of 5 KRHB buffer containing 38 mM glucose and 3 µg/ml of coelenterazine was then added to the cultured medium to acquire the luminescence video images.

Visualization by the luminescence imaging method is as follows. The luminescence signals produced from the cells as a result of luciferin-luciferase reaction of *Gaussia* luciferase with coelenterazine were taken at 37° C. using a model IX71 microscope (manufactured by Olympus Corporation) equipped with a thermostat incubator (manufactured by Tokai Hit Co.) and an EM-CCD camera (model C9100-13; 512×512 pixels, pixel size=16 µm; manufactured by Hamamatsu Photonics K. K.) in a dark box. The following objective lens with a high aperture number (NA) was used. UPLFLN 40× O (NA 1.30), ApoN 60× OTIRFM (NA 1.49) and PlanApo 100× 20 OTIRFM (NA 1.45) (manufactured by Olympus Corporation).

The luminescence video images were recorded onto the hard disk of a computer using an AQUACOSMOS software version 2.6 (manufactured by Hamamatsu Photonics K. K.). 25 The acquisition mode of the luminescence signal data using the AQUACOSMOS software above was used at 1×1 binning, fast scanning and photon-counting level=1. The luminescence video images acquired were processed and analyzed using the software above. In a few cases, the successive luminescence images were acquired using the "MaxTrace" method in the "sequential calculation" menu of the above software and displayed as picture images showing the maximum luminescence intensity.

The luminescence images acquired by the successive luminescence images were superimposed on the bright-field images to display the localization of luminescence signals in the individual cells. In order to analyze the time-lapse change of luminescence intensity change of luminescence intensity in video images, the mean luminescence intensity and maximum luminescence intensity within a particular luminescence region were calculated.

The transformant derived from MIN6 cell line transiently expressing the insulin-GLase fusion protein described above was analyzed by the luminescence imaging method, using an 45 EM-CCD camera-microscope device (objective lens x60; NA 1.49).

The luminescence signals of insulin-GLase fusion protein by exocytosis from the transformant derived from the MIN6 cell line cultured in a medium under low glucose conditions 50 (2 mM glucose) were little detected when observed in time resolution of 100-500 ms/frame.

After transfection, the insulin-GLase expression vector constructed in Example 1-1 above was cultured for 24 hours. This transiently transfected cell cluster derived from the Min6 55 cell line (FIG. 2, let: bright-field image) was stimulated by high glucose (20 mM glucose) medium containing coelenterazine. One minute after the onset of stimulation, luminescence video images were acquired for about 11.8 minutes at 100 ms/frame for 7000 frames.

After stimulation by 20 mM glucose, luminescence spots showing the secretion of the insulin-GLase fusion protein were frequently observed (FIG. 2, center: images of luminescence signals). By superimposing the bright-field images of the cells on the luminescence images, images of insulin secretion can be acquired. It is thus possible to identify the secretion sites in cells (FIG. 2, right).

34

Furthermore, editing of the successive high-speed luminescence images at 100 msec enables real-time imaging of insulin secretion (FIG. 3). As a result, editing of the successive composite images produced from the maximum luminescence intensities of all luminescence images revealed that the insulin-GLase fusion protein was secreted from the intercellular spaces of cell clusters of the transformant of MIN6 cells

The cells exhibiting a secretory luminescence are shown in the cell regions 1 to 3 at the center of FIG. 2, and time-lapse changes of the mean luminescence intensity in the cell regions 1 to 3 were calculated (FIG. 4).

The luminescence signals in the region 2 were detected in the narrow intercellular spaces where the cells interact with each other, and a few minutes after the high glucose stimulation, the luminescence intensity displayed a periodic pattern. The typical peaks of insulin-GLase secretion from the two cells in the regions 2 and 3 were analyzed, respectively (FIG. 4). These peaks represent the mass secretion of insulin-GLase generated by successive appearance of luminescence spots. These many luminescence signals generated by the mass secretion appeared successively around the edges of cells and then diffused in the region of intercellular spaces. This phenomenon was remarkable especially in the cells in the region 2. This enabled the dynamic analysis and quantification of insulin secretion at the same time.

In some other experiments, the cells associated with the close interaction between cells had a tendency to show pulsatile insulin secretion, as compared to the cells in the peripheral region of cell clusters (FIG. 5, FIG. 6).

In addition, the crowd of strong luminescent spots generated by secretion of the insulin-GLase fusion protein inside the cells was concentrated at a depth within 1 µm from the bottom of cell bodies. These data suggest that insulin secretion would be dependent on cell adhesion. These data also suggest that the formation of cell clusters which interact between cells would be important in a periodic secretion of insulin under physiological conditions.

In the secretion of the insulin-GLase fusion protein, the sporadic luminescence spots by exocytosis are easy to analyze the size and duration time of luminescence spots generated in a periodic pulsatile manner by secretion of the insulin-GLase fusion protein, as compared to the assembly of sporadic luminescence spots emitted by exocytosis. Many luminescence spots had a size not greater than 1 μ m and the duration time of the luminescence spots were between 0.1 and 1 second. These data suggest that luminescence signals by the secretion of each insulin-GLase would be generated by fusion of one cell granule (FIG. 7).

On the other hand, some luminescence signals continued over 1 to 30 seconds. These continuous spots were observed mainly on the bottom side of the cells (FIG. 8). It was first revealed that insulin secretion involved various modes of secretion.

Example 2-1

Glibenclamide, which is a sulfonylurea type hypoglycemic agent, promotes the secretion of endogenous insulin to lower blood glucose levels. This promotion of insulin secretion was examined using the insulin-GLase-expressed MIN6 cells to assess the effect. Specifically, the expression vector pcDNA3-hINS-GLuc prepared in EXAMPLE 1-1 was transformed onto MIN6 cells according to the method described in EXAMPLE 1-2 to prepare the insulin-GLase expression cells. After the MIN6 cell line was washed twice with 3 ml of phosphate buffered saline (PBS), 2 ml of low glucose-modi-

fied KRHB buffer solution containing 2 mM glucose for control (intact), 20 mM glucose and 10 mM glibenclamide (manufactured by Wako Pure Chemical), respectively, was added and an aliquot of 2 ml was taken every 0, 3, 4, 6, 8, 10, 15 and 30 minutes. The luciferase activity was assayed by the method described in EXAMPLE 1-2. As shown in FIG. 9, it was revealed that insulin secretion was promoted by treating with glibenclamide, indicating that drug efficacy can be assessed in this vector expression cell system.

Embodiment 3-1

Construction of Human Promatrix Metalloproteinase 2 (pro-hMMP-2)-*Gaussia* Luciferase (GLase) Fusion Protein

The expression vector pcDNA3-hMMP2-GLuc to express the fusion protein of pro-human matrix metalloproteinase 2 (pro-hMMP-2) having a signal peptide of human matrix metalloproteinase 2 (hMMP-2) and *Gaussia* luciferase (GLase) was constructed as follows.

The coding region of hMMP-2 proprotein was prepared by PCR (cycle conditions: 25 cycles; 15 sec/98° C., 15 sec/55° C., 2 mins/68° C.) using IMAGE cDNA clone 3161383 25 (cDNA clone prepared by the I.M.A.G.E. consortium). PCR was carried out using KOD-plus-DNA polymerase (manufactured by Toyobo Co., Ltd.) and primer sets of hMMP-2-P1 (5' ggc AAGCTT AGCCACC ATG GAG GCG CTA ATG GCC C 3'; BamHI recognition site underlined) (SEQ ID NO: 23) 30 and hMMP2-P2 (5' ggc GAATTC GCA GCC TAG CCA GTC GGA T 3'; EcoRI recognition site underlined) (SEQ ID NO: 24). The PCR fragment obtained was digested with recognition enzymes BamHI and EcoRI, which was the inserted into the BamHI/EcoRI site of the pcDNA3-GLuc-BE vector (described in FEBS Letters, 581, 4551-4556 (2007)) thereby to construct the expression vector pcDNA3hMMP2-GLuc. The protein expressed by the expression vector pcDNA3-hMMP2-GLuc obtained is the fusion protein 40 composed of the signal peptide sequence of hMMP-2, hMMP-2 and GLase.

Embodiment 3-2

Preparation of Transformant Using Expression Vector pcDNA3-hMMP2-GLuc

HeLa cells, which is the cell line derived from human uterine cervix carcinoma, were cultured in DMEM medium 50 (manufactured by Sigma) supplemented with 10% fetal calf serum (manufactured by Invitrogen).

The HeLa cell line was transfected by the expression vector pcDNA3-hMMP2-GLuc obtained in EMBODIMENT 3-1, using Fugene HD (manufactured by Roche). After the transfection, the cells were cultured at 37° C. for 24 hours in a CO_2 incubator.

Embodiment 3-3

Imaging of Extracellular Secretion of the Human Metalloproteinase 2 (hMMP-2)-GLase Fusion Protein

The fusion protein was cultured in a 35 mm uncoated 65 glass-bottom culture dish (manufactured by Asahi Glass Co., Ltd.) and imaging of extracellular secretion of the human

36

metalloproteinase 2 (hMMP-2)-GLase fusion protein was performed in a manner similar to EXAMPLE 1-3 described above

When GLase fused to hMMP-2 (hMMP-2-GLase fusion protein) was transiently expressed in HeLa cells for 24 hours, MMP-2-GLase in the medium showed about ½ luminescence intensity, as compared to that of GLase alone (FIG. 10). The HeLa cells transiently expressing the hMMP-2-GLase fusion protein and wild type hMMP-2 were analyzed by the immunofluorescence technique and the western blot technique, using anti-MMP-2 antibody and anti-GLase antibody. The hMMP-2-GLase fusion protein showed the localization as in wild type hMMP-2. These results demonstrate that the secretion pathway of hMMP-2-GLase is the same as in wild type hMMP-2.

On the other hand, the analysis of luminescence video images (500 msec/frame) in the transformed HeLa cells expressing the hMMP-2-GLase fusion protein demonstrated that the MMP-2 secretion showed cell polarity (FIG. 11). In the HeLa cells expressing the hMMP-2-GLase fusion protein, many bright luminescence spots were observed in the luminescence images immediately after addition of coelenterazine as a luminescent substrate, unlike the HeLa cells expressed by GLase alone. These luminescence spots first appeared stayed on the same place and then slowly diminished to the background level within a minute after addition of coelenterazine. By contrast, the presence of luminescence spots that maintained their luminescence indicates the hMMP-2-GLase fusion protein bound to the cell membrane surface. These continuing luminescence spots move toward the front of a migrating cell to give an uneven distribution. MMP-2 is known to bind integrin or MT1-MMP (MMP14) (this is an activator of MMP-2) on the cell surface. These continuing luminescence spots of the MMP-2-GLase fusion protein show the localization of MMP-2 on the cell surface, bound to the protein described above.

The luminescence spots showing exocytosis of the hMMP-2-GLase fusion protein, which transiently appeared, were observed until or after the continuing luminescence spots disappeared. Under the same imaging conditions (500 msec/frame, 40× objective lens; FIG. 12), the luminescence spots of hMMP-2-GLase transiently appeared were maintained in narrow regions and slowly diffused over a few seconds, unlike the luminescence spots of non-fused GLase alone.

The luminescence spots by secretion of the hMMP-2-GLase fusion protein emerged chiefly around the leading edge of the cells. Successive secretion of the hMMP-2-GLase fusion protein over a few seconds was sometimes observed along the leading edge (FIG. 12). The MMP-2 protease activity may be required to degrade cell adhesion protein and lift up ruffling membranes, and successive secretion of hMMP-2-GLase may be associated with the formation and migration of ruffling membranes at the leading edge. The luminescence signals by hMMP-2-GLase secretion were not stimulated by increased K⁺ concentrations and clearly were not affected by intracellular ions

After the continuous secretory luminescence disappeared (FIG. 13), the maximum luminescence signals (FIG. 14, left) generated from exocytosis of the hMMP-2-GLase fusion protein were superimposed on the bright-field image to produce the composite image (100 frames for about 50 secs.; FIG. 14, right). Localization and frequency of exocytosis phenomenon of the hMMP-2-GLase fusion protein were estimated. This composite image clearly indicates that MMP-2 shows a polar distribution around the leading edge of a migrating cell. The distribution of luminescence spots by the exocytosis was closer to the leading edge, as compared to the distribution of

continuing luminescence spots observed at an early stage (FIG. 14). The luminescence video images and composite image (FIG. 14, right) also shows the secretion of the hMMP-2-GLase fusion protein from a migrating cell. These data suggest that hot spots of MMP-2 secretion by the exocytosis phenomenon and punctate spots of MMP-2 remained on the cell surface are localized on the discrete minimal region of plasma membrane.

Next, the frequency of exocytosis in the minimal region of cells was estimated. In FIG. 13, the composite image of the maximum luminescence intensity in the four regions circled in the composite image in FIG. 13 showed several luminescence peaks in each region (FIG. 15). The marked luminescence peaks (designated by the arrow in FIG. 14) in the regions 1 to 4 were measured and found to be 3, 4, 1 and 2, respectively. These numbers represent the number of marked exocytosis that occurred at least during this period (100 frames, about 50 seconds). These data indicate that the exocytosis of the hMMP-2-GLase fusion protein is repeated in the limited minimal region on the cell surface of a migrating cell.

Next, imaging was performed using a 100x objective lens (number of aperture NA, 1.45). Immediately after acquisition of the luminescence video images using a 40x objective lens, the luminescence video images associated with hMMP-2-GLase of the same migrating cell were acquired at 500 ms/frame (200 frames for about 100 seconds). The luminescence spots by each secretion of the hMMP-2-GLase fusion protein at the leading edge were more clearly observed from the luminescence images acquired using a high 100x objective lens. The luminescence spots (FIG. 16, spots 1 to 6) of the luminescence video images in one frame designated by the arrows were determined to be the signals generated by the exocytosis phenomenon, based on the images. The spots 2 to 4 newly emerged in this frame. In spots 2 and 3, the luminescence intensity of the luminescence spots by exocytosis diminished to the background level in 2 to 3 seconds (FIG. 35 17). Some luminescence spots such as spot 2 which repeatedly appeared suggest hMMP-2 secretion due to continuous membrane fusion of exocytotic secretory granules (FIG. 17).

Most of the luminescence spots from the hMMP-2-GLase fusion protein are less than 1 µm, like these spots. The diameter of the luminescence spot from one exocytotic granule was measured to be 0.1 to 0.4 µm by TIRF and the two-photon imaging method (Science, 297, 1349-1352 (2002)). Accordingly, these luminescence spots are generated either by secretion of MMP-2-GLase fusion protein derived from one exocytotic secretory granule or by continuous fusion of secretory vesicles in the minimal region. In the composite image of the maximum luminescence intensity in the video images, distribution of the exocytosis spots at the leading edge can be analyzed with a high resolution. The number of the luminescence spots transiently appeared in the region 1 and region 2 was calculated to be 70 and 20, respectively (FIG. 16, lower right).

The luminescence spots by secretion of the hMMP-2-GLase fusion protein were concentrated at a depth within 1 55 µm from the bottom side of cells. This suggests that hMMP-2 would be secreted mainly from the bottom side of the cell surface.

These data suggest that the fusion of exocytotic secretory granules including MMP-2 would take place at the leading edge for a short period of time to disrupt cell adhesion and extracellular matrix.

INDUSTRIAL APPLICABILITY

The screening method of the present invention is useful for screening drugs such as insulin secretagogues having an insu-

38

lin secretagogue activity with minimized side effects (hypoglycemia induction, etc.). The transformant in which a polynucleotide encoding the fusion protein used for the screening method is introduced, the screening kit comprising the transformant, etc. are also useful for screening excellent drugs.

BRIEF DESCRIPTION OF THE SEQUENCES

SEQ ED NO: 1 Nucleotide sequence of the polynucleotide encoding *Gaussia* luciferase (GLase)

SEQ ID NO: 2 Amino acid sequence of *Gaussia* luciferase SEQ ID NO: 3 Nucleotide sequence of the polynucleotide encoding human proinsulin

SEQ ID NO: 4 Amino acid sequence of human proinsulin SEQ ID NO: 5 Nucleotide sequence of the polynucleotide encoding the signal peptide of human preproinsulin

SEQ ID NO: 6 Amino acid sequence of the signal peptide of human preproinsulin SEQ ID NO: 7 Nucleotide sequence of the polynucleotide encoding *Gaussia* luciferase (GLase (K18-D185)) lacking the signal sequence, used in Example 1-1 and EMBODIMENT 3-1

SEQ ID NO: 8 Amino acid sequence of the polypeptide (GLase (K18-D185)) encoded by the polynucleotide consisting of the nucleotide sequence represented by SEQ ID NO: 7

SEQ ID NO: 9 Nucleotide sequence of the polynucleotide encoding human preproinsulin used in EXAMPLE 1-1

SEQ ID NO: 10 Amino acid sequence of the polypeptide (human preproinsulin) encoded by the polynucleotide consisting of the nucleotide sequence represented by SEQ ID NO: 9

SEQ ID NO: 11 Nucleotide sequence of the polynucleotide encoding the fusion protein of human preproinsulin and *Gaussia* luciferase (GLase (K18-D185)) lacking the signal sequence, used in EXAMPLE 1-1

SEQ ID NO: 12 Amino acid sequence of the polypeptide encoded by the polynucleotide consisting of the nucleotide sequence represented by SEQ ID NO: 11

SEQ ID NO: 13 Nucleotide sequence of the polynucleotide encoding human MMP-2

SEQ ID NO: 14 Amino acid sequence of human MMP-2

SEQ ID NO: 15 Amino acid sequence of the polynucleotide encoding the signal sequence of human pro-MMP-2

SEQ ID NO: 16 Amino acid sequence of the signal sequence of human pro-MMP-2

SEQ ID NO: 17 Nucleotide sequence of the polynucleotide encoding human pro-MMP-2 used in EMBODIMENT 3-1

SEQ ID NO: 18 Amino acid sequence of the polypeptide (human pro-MMP-2) encoded by the polynucleotide represented by SEQ ID NO: 17

SEQ ID NO: 19 Nucleotide sequence of the polynucleotide encoding the fusion protein of human pro-MMP-2 and *Gaussia* luciferase (GLase (K18-D185)) lacking the signal sequence, used in EMBODIMENT 3-1

SEQ ID NO: 20 Amino acid sequence of the polypeptide encoded by the polynucleotide represented by SEQ ID NO: 19

60 SEQ ID NO: 21 Nucleotide sequence of the primer used in EXAMPLE 1-1

SEQ ID NO: 22 Nucleotide sequence of the primer used in EXAMPLE 1-1

SEQ ID NO: 23 Nucleotide sequence of the primer used in EMBODIMENT 3-1

SEQ ID NO: 24 Nucleotide sequence of the primer used in EMBODIMENT 3-1

SEQUENCE LISTING

<160	JN <	JMBEF	OF	SEQ	ID 1	10S:	24									
<211 <212 <213 <220 <221	L> LE 2> TY 3> OF 0> FE L> NF	EQ II ENGTH PE: RGANI EATUF AME/F	I: 59 DNA SM: RE: KEY:	Gaus CDS		-	nceps	3								
		QUEN			,	,,,										
		gtc Val														48
		ccc Pro														96
		ttc Phe 35														144
		aag Lys														192
		gct Ala		_				_	_		_	_				240
_	_	acg Thr		_	_	_	_					_	_			288
		ggc Gly														336
		att Ile 115														384
		atc Ile														432
		gly ggg		_			_	_		_	_		_	_		480
_	_	caa Gln	_	_				_	_	_		_		_		528
		atc Ile														555
<211 <212	L> LE 2> TY	EQ II ENGTH PE: RGANI	H: 18 PRT	35	ssia	prin	nceps	3								
		EQUEN			·	-										
Met 1	Gly	Val	Lys	Val 5	Leu	Phe	Ala	Leu	Ile 10	Cys	Ile	Ala	Val	Ala 15	Glu	
Ala	Lys	Pro	Thr 20	Glu	Asn	Asn	Glu	Asp 25	Phe	Asn	Ile	Val	Ala 30	Val	Ala	
Ser	Asn	Phe 35	Ala	Thr	Thr	Asp	Leu 40	Asp	Ala	Asp	Arg	Gly 45	Lys	Leu	Pro	

-continued

```
Gly Lys Lys Leu Pro Leu Glu Val Leu Lys Glu Met Glu Ala Asn Ala
                         55
Arg Lys Ala Gly Cys Thr Arg Gly Cys Leu Ile Cys Leu Ser His Ile
Lys Cys Thr Pro Lys Met Lys Lys Phe Ile Pro Gly Arg Cys His Thr
Tyr Glu Gly Asp Lys Glu Ser Ala Gln Gly Gly Ile Gly Glu Ala Ile
Val Asp Ile Pro Glu Ile Pro Gly Phe Lys Asp Leu Glu Pro Met Glu
Gln Phe Ile Ala Gln Val Asp Leu Cys Val Asp Cys Thr Thr Gly Cys
Leu Lys Gly Leu Ala Asn Val Gln Cys Ser Asp Leu Leu Lys Lys Trp
Leu Pro Gln Arg Cys Ala Thr Phe Ala Ser Lys Ile Gln Gly Gln Val
Asp Lys Ile Lys Gly Ala Gly Gly Asp
180 185
<210> SEO ID NO 3
<211> LENGTH: 258
<212> TYPE: DNA
<213 > ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (1)..(258)
<400> SEOUENCE: 3
ttt gtg aac caa cac ctg tgc ggc tca cac ctg gtg gaa gct ctc tac
                                                                          48
Phe Val Asn Gln His Leu Cys Gly Ser His Leu Val Glu Ala Leu Tyr
     5
                                     10
cta gtg tgc ggg gaa cga ggc ttc ttc tac aca ccc aag acc cgc cgg
                                                                          96
Leu Val Cys Gly Glu Arg Gly Phe Phe Tyr Thr Pro Lys Thr Arg Arg
                                 25
gag gca gag gac ctg cag gtg ggg cag gtg gag ctg ggc ggc cct
Glu Ala Glu Asp Leu Gln Val Gly Gln Val Glu Leu Gly Gly Gly Pro
                                                                         144
        35
                             40
ggt gca ggc agc ctg cag ccc ttg gcc ctg gag ggg tcc ctg cag aag
                                                                         192
Gly Ala Gly Ser Leu Gln Pro Leu Ala Leu Glu Gly Ser Leu Gln Lys
                         55
cgt ggc att gtg gaa caa tgc tgt acc agc atc tgc tcc ctc tac cag
Arg Gly Ile Val Glu Gln Cys Cys Thr Ser Ile Cys Ser Leu Tyr Gln
                    70
ctg gag aac tac tgc aac
Leu Glu Asn Tyr Cys Asn
<210> SEQ ID NO 4
<211> LENGTH: 86
<212> TYPE: PRT
<213 > ORGANISM: Homo sapiens
<400> SEQUENCE: 4
Phe Val Asn Gln His Leu Cys Gly Ser His Leu Val Glu Ala Leu Tyr
                       10
Leu Val Cys Gly Glu Arg Gly Phe Phe Tyr Thr Pro Lys Thr Arg Arg
                     25
Glu Ala Glu Asp Leu Gln Val Gly Gln Val Glu Leu Gly Gly Pro
```

35 40

-continued

```
Gly Ala Gly Ser Leu Gln Pro Leu Ala Leu Glu Gly Ser Leu Gln Lys
    50
                          55
                                                60
Arg Gly Ile Val Glu Gln Cys Cys Thr Ser Ile Cys Ser Leu Tyr Gln
                     70
                                            75
Leu Glu Asn Tyr Cys Asn
<210> SEQ ID NO 5
<211> LENGTH: 72
<212> TYPE: DNA
<213 > ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (1)..(72)
<400> SEQUENCE: 5
atg gcc ctg tgg atg cgc ctc ctg ccc ctg ctg gcg ctg ctg gcc ctc Met Ala Leu Trp Met Arg Leu Leu Pro Leu Leu Ala Leu Leu Ala Leu
                                                                             48
tgg gga cct gac cca gcc gca gcc
Trp Gly Pro Asp Pro Ala Ala Ala
                                                                             72
           20
<210> SEQ ID NO 6
<211> LENGTH: 24
<212> TYPE: PRT
<213 > ORGANISM: Homo sapiens
<400> SEQUENCE: 6
Met Ala Leu Trp Met Arg Leu Leu Pro Leu Leu Ala Leu Leu Ala Leu
                                       10
Trp Gly Pro Asp Pro Ala Ala Ala
            20
<210> SEQ ID NO 7
<211> LENGTH: 504
<212> TYPE: DNA
<213> ORGANISM: Gaussia princeps
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (1)..(504)
<400> SEQUENCE: 7
aag ccc acc gag aac aac gaa gac ttc aac atc gtg gcc gtg gcc agc
                                                                             48
Lys Pro Thr Glu Asn Asn Glu Asp Phe Asn Ile Val Ala Val Ala Ser
aac ttc gcg acc acg gat ctc gat gct gac cgc ggg aag ttg ccc ggc
Asn Phe Ala Thr Thr Asp Leu Asp Ala Asp Arg Gly Lys Leu Pro Gly
aag aag ctg ccg ctg gag gtg ctc aaa gag atg gaa gcc aat gcc cgg
Lys Lys Leu Pro Leu Glu Val Leu Lys Glu Met Glu Ala Asn Ala Arg
                                                                            192
aaa get gge tge ace agg gge tgt etg ate tge etg tee eac ate aag
Lys Ala Gly Cys Thr Arg Gly Cys Leu Ile Cys Leu Ser His Ile Lys
                         55
                                                60
tgc acg ccc aag atg aag aag ttc atc cca gga cgc tgc cac acc tac
                                                                            240
Cys Thr Pro Lys Met Lys Lys Phe Ile Pro Gly Arg Cys His Thr Tyr
65
                     70
gaa ggc gac aaa gag tcc gca cag ggc ggc ata ggc gag gcg atc gtc
Glu Gly Asp Lys Glu Ser Ala Gln Gly Gly Ile Gly Glu Ala Ile Val
                 85
                                       90
gac att cct gag att cct ggg ttc aag gac ttg gag ccc atg gag cag
Asp Ile Pro Glu Ile Pro Gly Phe Lys Asp Leu Glu Pro Met Glu Gln
```

									US	9,1	81,	318	B2	
				45										46
									-	con	tin [.]	ued		
	100					105					110			
ttc atc go Phe Ile Al 11	a Gln													384
aaa ggg ct Lys Gly Le	t gcc			Gln	tgt				Leu	aag				432
130 ccg caa cg Pro Gln Ar			Thr					Ile					Asp	480
145 aag atc aa								155					160	504
Lys Ile Ly	s Gly	Ala 165	Gly	Gly	Asp									
<210> SEQ <211> LENG <212> TYPE <213> ORGA	TH: 1 : PRT	68	ssia	pri	ncepi	s								
<400> SEQU	ENCE:	8												
Lys Pro Th	r Glu	Asn 5	Asn	Glu	Asp	Phe	Asn 10	Ile	Val	Ala	Val	Ala 15	Ser	
Asn Phe Al	a Thr 20	Thr	Asp	Leu	Asp	Ala 25	Asp	Arg	Gly	ГÀа	Leu 30	Pro	Gly	
Lys Lys Le		Leu	Glu	Val	Leu 40	ГЛа	Glu	Met	Glu	Ala 45	Asn	Ala	Arg	
Lys Ala Gl 50	у Сув	Thr	Arg	Gly 55	Cys	Leu	Ile	Cys	Leu 60	Ser	His	Ile	Lys	
Cys Thr Pr 65	o Lys	Met	Lys 70	Lys	Phe	Ile	Pro	Gly 75	Arg	Cys	His	Thr	Tyr 80	
Glu Gly As	p Lys	Glu 85	Ser	Ala	Gln	Gly	Gly 90	Ile	Gly	Glu	Ala	Ile 95	Val	
Asp Ile Pr	o Glu 100	Ile	Pro	Gly	Phe	Lys 105	Asp	Leu	Glu	Pro	Met 110	Glu	Gln	
Phe Ile Al 11		Val	Asp	Leu	Сув 120	Val	Asp	CAa	Thr	Thr 125	Gly	CÀa	Leu	
Lys Gly Le 130				135					140					
Pro Gln Ar 145			150			Ser	Lys	Ile 155	Gln	Gly	Gln	Val	Asp 160	
Lys Ile Ly	s Gly	Ala 165	Gly	Gly	Asp									
<210 > SEQ <211 > LENG <212 > TYPE <213 > ORGA <220 > FEAT <221 > NAME	TH: 3 : DNA NISM: URE: /KEY:	30 Hom CDS			s									
<222> LOCA			(3.	∍ ∪)										
atg gcc ct Met Ala Le 1														48
tgg gga co Trp Gly Pr														96

tca cac ctg gtg gaa gct ctc tac cta gtg tgc ggg gaa cga ggc ttc

-continued

Ser Nis Lew Val Glu Als Lew Tyr Lew Val Cye Gly Glu Arg Gly Pice	_												con	tını —	ued 		
Phe Typ Thr Pro Lys Thr Ang Arg Gil Als Gil Asp Leu Gil Val Gily 50	Ser	His		Val	Glu	Ala	Leu	_	Leu	Val	Сув	Gly		Arg	Gly	Phe	
01n Vai Glu Leu Gly Gly Gly Gly Pro Gly Ala Gly Ser Leu Gln Pro Lou 55 70 80 90 90 90 90 90 90 90 90 90 90 90 90 90		Tyr					Arg					Asp					192
Ala Leu Glu Gly Ser Leu Gln Lye Arg Gly 11e Val Glu Gln Cye Cye 85 acc agc atc tgc tcc ctc tac cag ctg gag aac tac tgc aac Thr Ser 11e Cye Ser Leu Tyr Gln Leu Glu Asn Tyr Cye Asn 100	Gln					Gly					Gly					Leu	240
Thr Ser Ile Cys Ser Leu Tyr Chi Leu Glu Asn Tyr Cys Asn 110 Callo SEQ ID NO 10 Callo Type: PRT Callo Rev Cys Cys Ser Leu Type: PRT Callo Rev Cys Cys Ser Leu Type: PRT Callo Rev Cys Cys Ser His Leu Val Glu Ala Leu Type: Leu Leu Leu Ala Leu Leu Ala Leu Cys Gly 20 Callo Rev Cys Cys Ser His Leu Val Glu Ala Leu Tyr Leu Val Cys Gly Glu Arg Gly Phe 35 Callo Rev Cys Cys Ser His Leu Val Glu Ala Leu Tyr Leu Val Cys Gly Glu Arg Gly Phe 35 Callo Rev Cys Cys Ser His Leu Val Glu Leu Gly Gly Pro Gly Ala Gly Ser Leu Gln Pro Leu Glu Cys Cys Ser Cys Ser Leu Glu Leu Gly Gly Pro Gly Ala Gly Ser Leu Gln Pro Leu Glu Cys Cys Ser Leu Glu Leu Gly Gly Gly Pro Gly Ala Gly Ser Leu Gln Pro Leu Gys Cys Ser Leu Glu Cys Cys Ser Cys Cys Cys Ser Cys Cys Cys Ser Cys Cys Ser Cys Cys Cys Ser Cys Cys Cys Ser Cys Cys Cys Ser Cys Cys Cys Cys Ser Cys Cys Cys Cys Ser Cys					Ser					Gly					Cys		288
*211> LENOTH: 110 *212> TOPE: PRT *213> ORGANISM: Homo sapiens *400> SEQUENCE: 10 Met Ala Leu Trp Met Arg Leu Leu Pro Leu Leu Ala Leu Leu Ala Leu 1				Cys	Ser				Leu					Asn			330
Met Ala Leu Trp Met Arg Leu Leu Pro Leu Leu Ala Leu Leu Ala Leu Leu Ala Leu Trp Gly Pro App Pro Ala Ala Ala Phe Val Asn Gln His Leu Cys Gly 20 Ser His Leu Val Glu Ala Leu Tyr Leu Val Cys Gly Glu Arg Gly Phe 35 Ala Leu Gly Gly Gly Fro Gly Ala Glu Ala Glu App Leu Gln Val Gly 50 Gln Val Glu Leu Gly Gly Gly Pro Gly Ala Gly Ser Leu Gln Pro Leu 65 Ala Leu Glu Gly Ser Leu Gln Lys Arg Gly Ile Val Glu Gly Cys 95 Thr Ser Ile Cys Ser Leu Tyr Gln Leu Glu Asn Tyr Cys Asn 100	<21 <21	1> Ll 2> T	ENGTI YPE :	H: 1	10	o saj	piens	g									
1						Arq	Leu	Leu	Pro	Leu	Leu	Ala	Leu	Leu	Ala	Leu	
Ser His Leu Val Glu Ala Leu Tyr Leu Val Cys Gly Glu Arg Gly Phe 35 Ala Leu Glu Gly Gly Gly Pro Gly Ala Glu Asp Leu Gln Val Gly 55 Cln Val Glu Leu Gly Gly Gly Pro Gly Ala Gly Ser Leu Gln Pro Leu 65 Ala Leu Glu Gly Ser Leu Gln Lys Arg Gly Ile Val Glu Gln Cys 85 Thr Ser Ile Cys Ser Leu Tyr Gln Leu Glu Asn Tyr Cys Asn 100 100 100 210> SEQ ID NO 11 211- LENGTH: 840 212- YPE' DNA 212- YPE' DNA 212- YPE' DNA 212- YPE' CNS 222- LOCATION: (1) (840) 4400> SEQUENCE: 11 atg gcc ctg tgg atg cgc ctc ctg ccc ctg ctg gcg ctg ctg gcg ctc Met Ala Leu Trp Met Arg Leu Leu Pro Leu Leu Ala Leu 1	1			Ī	5	_				10					15		
25	Ī	_		20					25					30		-	
50 55 60 60 60 60 60 60 60 60 60 60 60 60 60			35					40			-	-	45		•		
Ala Leu Glu Gly Ser Leu Gln Lys Arg Gly Ile Val Glu Gln Cys Cys 95 Thr Ser Ile Cys Ser Leu Tyr Gln Leu Glu Asn Tyr Cys Asn 110 <pre></pre>	Phe		Thr	Pro	ГÀа	Thr	_	Arg	Glu	Ala	Glu	_	Leu	GIn	Val	GIĄ	
Thr Ser Ile Cys Ser Leu Tyr Gln Leu Glu Asn Tyr Cys Asn 100 <pre></pre>		ı Val	Glu	Leu	Gly		Gly	Pro	Gly	Ala	_	Ser	Leu	Gln	Pro		
<pre>2210 > SEQ ID NO 11 2211 > LENGTH: 840 2212 > TYPE: DNA 2213 > ORGANISM: Artificial 2220 > FEATURE: 2223 > OTHER INFORMATION: fusion protein 2201 > FEATURE: 2213 > NAME/KEY: CDS 2222 > LOCATION: (1) (840) 400 > SEQUENCE: 11 atg gcc ctg tgg atg cgc ctc ctg ccc ctg ctg gcg ctg ctg gcc ctc Met Ala Leu Trp Met Arg Leu Leu Pro Leu Leu Ala Leu Leu Ala Leu 1</pre>	Ala	Leu	Glu	Gly		Leu	Gln	Lys	Arg	_	Ile	Val	Glu	Gln	_	CAa	
<pre></pre>	Thr	Ser	Ile	_	Ser	Leu	Tyr	Gln		Glu	Asn	Tyr	Сув				
atg gcc ctg tgg atg cgc ctg ctg gcc ctc ctg ccc ctg ctg c	<21 <21 <22 <22 <22 <22 <22	1 > L1 2 > T 3 > O1 0 > F1 3 > O' 0 > F1 1 > NA	ENGTI YPE: RGAN: EATUI THER EATUI AME/I	H: 8 DNA ISM: RE: INF RE: KEY:	Art: ORMA' CDS	TION	: fu:	sion	prot	cein							
Met Ala Leu Trp Met Arg Leu Leu Pro Leu Ala Ala Leu Ala Ala <td< td=""><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></td<>																	
Trp Gly Pro Asp Pro Ala Ala Ala Phe Val Asn Gln His Leu Cys Gly 30 tca cac ctg gtg gaa gct ctc tac cta gtg tgc ggg gaa cga ggc ttc Ser His Leu Val Glu Ala Leu Tyr Leu Val Cys Gly Glu Arg Gly Phe 35 ttc tac aca ccc aag acc cgc cgg gag gca gag gca ctg cag gtg ggg 192 ttc tac aca ccc aag acc cgc cgg gag gca gag gca ctg cag gtg ggg 192 Phe Tyr Thr Pro Lys Thr Arg Arg Glu Ala Glu Ala Glu Asp Leu Gln Val Gly 50 cag gtg gag ctg ggc ggc ggc cct ggt gca ggc agc ctg cag ccc ttg 240	Met				Met					Leu					Ala		
Ser His Leu Val Glu Ala Leu Tyr Leu Val Cys Gly Glu Arg Gly Phe 35 ttc tac aca ccc aag acc cgc cgg gag gca gag gca ctg cag gtg ggg 192 Phe Tyr Thr Pro Lys Thr Arg Arg Glu Ala Glu Asp Leu Gln Val Gly 50 cag gtg gag ctg ggc ggc ggc cct ggt gca ggc agc ctg cag ccc ttg 240				Āsp					Phe					Leu			96
Phe Tyr Thr Pro Lys Thr Arg Arg Glu Ala Glu Asp Leu Gln Val Gly 50 55 60 cag gtg gag ctg ggc ggc ggc cct ggt gca ggc agc ctg cag ccc ttg 240			Leu					Tyr					Glu				144
		Tyr					Arg					Asp					192
																	240

-continued

		-continued	
65	70 75	5 8	0
	ctg cag aag cgt ggc at Leu Gln Lys Arg Gly Il 90		=
	ctc tac cag ctg gag aa Leu Tyr Gln Leu Glu As 105		
	aac gaa gac ttc aac at Asn Glu Asp Phe Asn Il 120		
	gat ete gat get gae eg Asp Leu Asp Ala Asp Ar 135		
Lys Lys Leu Pro Leu	gag gtg ctc aaa gag at Glu Val Leu Lys Glu Me 150 15	et Glu Ala Asn Ala A	
	agg ggc tgt ctg atc tg Arg Gly Cys Leu Ile Cy 170	=	
	aag aag ttc atc cca gg Lys Lys Phe Ile Pro Gl 185		
	tcc gca cag ggc ggc at Ser Ala Gln Gly Gly Il 200		
	cct ggg ttc aag gac tt Pro Gly Phe Lys Asp Le 215		
Phe Ile Ala Gln Val	gat ctg tgt gtg gac tg Asp Leu Cys Val Asp Cy 230 23	s Thr Thr Gly Cys L	
	gtg cag tgt tct gac ct Val Gln Cys Ser Asp Le 250		
	acc ttt gcc agc aag at Thr Phe Ala Ser Lys Il 265		
aag atc aag ggg gcc Lys Ile Lys Gly Ala 275			840
<210> SEQ ID NO 12 <211> LENGTH: 280 <212> TYPE: PRT <213> ORGANISM: Arti <220> FEATURE: <223> OTHER INFORMAT	ficial TON: Synthetic Constru	ıct	
<400> SEQUENCE: 12			
Met Ala Leu Trp Met 1 5	Arg Leu Leu Pro Leu Le 10	eu Ala Leu Leu Ala L 15	eu
Trp Gly Pro Asp Pro 20	Ala Ala Ala Phe Val As 25	sn Gln His Leu Cys G 30	ily
Ser His Leu Val Glu 35	Ala Leu Tyr Leu Val Cy 40	vs Gly Glu Arg Gly P 45	he
Phe Tyr Thr Pro Lys 50	Thr Arg Arg Glu Ala Gl 55	lu Asp Leu Gln Val G 60	ily

Gln Val Glu Leu Gly Gly Gly Pro Gly Ala Gly Ser Leu Gln Pro Leu 65 70707575 Ro Leu Gly Ro Ro

-continued

Ala	Leu	Glu	Gly	Ser 85	Leu	Gln	Lys	Arg	Gly 90	Ile	Val	Glu	Gln	Сув 95	Cys	
Thr	Ser	Ile	Cys	Ser	Leu	Tyr	Gln	Leu 105	Glu	Asn	Tyr	Cys	Asn 110	Glu	Phe	
Lys	Pro	Thr 115	Glu	Asn	Asn	Glu	Asp 120		Asn	Ile	Val	Ala 125	Val	Ala	Ser	
Asn	Phe 130	Ala	Thr	Thr	Asp	Leu 135	Asp	Ala	Asp	Arg	Gly 140	Lys	Leu	Pro	Gly	
Lys 145	-	Leu	Pro	Leu	Glu 150	Val	Leu	Lys	Glu	Met 155	Glu	Ala	Asn	Ala	Arg 160	
Lys	Ala	Gly	Cys	Thr 165	Arg	Gly	Cys	Leu	Ile 170		Leu	Ser	His	Ile 175	Lys	
Càa	Thr	Pro	Lys 180	Met	Lys	Lys	Phe	Ile 185	Pro	Gly	Arg	Càa	His 190	Thr	Tyr	
Glu	Gly	Asp 195		Glu	Ser	Ala	Gln 200		Gly	Ile	Gly	Glu 205	Ala	Ile	Val	
Asp	Ile 210	Pro	Glu	Ile	Pro	Gly 215	Phe	Lys	Asp	Leu	Glu 220	Pro	Met	Glu	Gln	
Phe 225	Ile	Ala	Gln	Val	Asp 230	Leu	Cya	Val	Asp	Сув 235	Thr	Thr	Gly	CÀa	Leu 240	
ГЛа	Gly	Leu	Ala	Asn 245	Val	Gln	Cya	Ser	Asp 250	Leu	Leu	ГЛа	ГЛа	Trp 255	Leu	
Pro	Gln	Arg	Cys 260	Ala	Thr	Phe	Ala	Ser 265		Ile	Gln	Gly	Gln 270	Val	Asp	
Lys	Ile	Lys 275	Gly	Ala	Gly	Gly	Asp 280									
<211 <212 <213 <220 <221	D> FE L> NA	ENGTH (PE: RGAN] EATUH AME/H	H: 10 DNA ISM: RE: KEY:	553 Homo	ວ sa <u>r</u> (10		3									
<211 <212 <213 <220 <221 <222	L> LE 2> T\ 3> OF D> FE L> NF	ENGTH (PE: RGANI EATUR AME / R DCATI	H: 16 DNA ISM: RE: CEY: ION:	Homo CDS (1)			3									
<211 <212 <213 <220 <221 <222 <400 tac	L> LE 2> T) 3> OF 0> FE L> NF 2> LO 0> SE	ENGTH (PE: RGANI EATUR AME/H DCATI EQUEN	H: 16 DNA ISM: RE: REY: ION: ICE:	DS (1) 13 cct		353) aag	ccc									48
<211 <212 <213 <220 <221 <222 <400 tac Tyr 1 tac	L> LE 2> TY 3> OF 0> FE L> NA 2> LO 0> SE aac Asn	ENGTH (PE: (GAN) EATUH AME/H CCATI EQUEN ttc Phe atc	H: 16 DNA ISM: RE: CEY: ION: Ttc Phe att	Homo CDS (1) 13 cct Pro 5	(16	aag Lys aca	ccc Pro	Lys gat	Trp 10 ctg	Asp gac	Lys	Asn gag	Gln	Ile 15 gtg	Thr gat	48
<211 <212 <213 <220 <222 <400 tac Tyr tac Tyr	L> LE 2> TY 3> OF 0> FE L> NA 2> LO 0> SE aac Asn agg Arg	ENGTH (PE: RGANI) EATUH AME/H DCATI EQUEN ttc Phe atc Ile	H: 16 DNA ISM: SE: CEY: ON: TCE: ttc Phe att Ile 20 gct	Homo CDS (1) 13 cct Pro 5 ggc Gly	cgc Arg	aag Lys aca Thr	ccc Pro cct Pro	Lys gat Asp 25 gtc	Trp 10 ctg Leu	Asp gac Asp	Lys cca Pro gat	Asn gag Glu gtg	Gln aca Thr 30 acc	Ile 15 gtg Val	Thr gat Asp	
<211 <212 <213 <222 <222 <400 tac Tyr 1 tac Tyr gat Asp	1> LE 2> T) 3> OF 0> FF 1> NAM 1> NAM 2> LC Asn agg Arg gcc Ala	ENGTH (PE: (PE: (PE: (PE: (PE: (PE: (PE: (PE:	H: 16 DNA ISM: RE: RE: REY: ION: ICE: ttc Phe att 120 gct Ala	CDS (1) 13 cct Pro 5 ggc Gly cgt Arg	cgc Arg tac Tyr	aag Lys aca Thr ttc Phe	ccc Pro cct Pro caa Gln 40	gat Asp 25 gtc Val	Trp 10 ctg Leu tgg Trp	Asp gac Asp agc Ser	cca Pro gat Asp	Asn gag Glu gtg Val 45 atg	Gln aca Thr 30 acc Thr	Ile 15 gtg Val cca Pro	Thr gat Asp ctg Leu	96
<211 <212 <213 <220 <221 <400 tac Tyr tac Tyr gat Asp	l> LE 2> TY 3> OF	ENGTH (PE: (PE: (PE: (PE: (PE: (PE: (PE: (PE:	H: 16 DNA SM: RE: CON: UCE: ttc Phe attt Ile 20 gct Ala cga Arg	Homo CDS (1) 13 cet Pro 5 ggc Gly cgt Arg	cgc Arg tac Tyr gcc Ala	aag Lys aca Thr ttc Phe gat Asp 55	ccc Pro cct Pro caa Gln 40 gga Gly	gat Asp 25 gtc Val gag Glu	Trp 10 ctg Leu tgg Trp gca Ala	Asp gac Asp agc Ser gac Asp	cca Pro gat Asp atc Ile 60	Asn gag Glu gtg Val 45 atg Met	aca Thr 30 acc Thr	Ile 15 gtg Val cca Pro aac Asn	Thr gat Asp ctg Leu ttt Phe	96 144
<211 <211 <211 <211 <211 <221 <222 <400 tac Tyr tac Tyr gat Asp cgg Arg ggc Gly 65 ctc	L> LE 2> TY 3> OF 1> NA 2> LC Asn agg Arg gcc Ala ttt Phe 50 cgc Arg	ENGTH (PE: SEATURE AND ADDRESS	H: 16 DNA: ISE: ISE: ISE: ISE: ISE: ISE: ISE: ISE	Homo CDS (1) 13 cct Pro 5 ggc Gly cgt Arg atc Ile cat His	cgc Arg tac Tyr gcc Ala cat His	aag Lys aca Thr ttc Phe gat Asp 55 gat Asp	cct Pro cct Pro caa Gln 40 gga Gly	gat Asp 25 gtc Val gag Glu tac Tyr	Trp 10 ctg Leu tgg Trp gca Ala ccc Pro	gac Asp agc Ser gac Asp ttt Phe 75	cca Pro gat Asp atc Ile 60 gac Asp	Asn gag Glu gtg Val 45 atg Met ggt Gly	Gln aca Thr 30 acc Thr atc Ile aag Lys	Ile 15 gtg Val cca Pro aac Asn gac	gat Asp ctg Leu ttt Phe gga Gly 80 tcc	96 144 192
<211: <211: <212: <212: <222: <400 tac Tyr tac Tyr gat Asp cgg Arg ggc Gly 65 ctc Leu cat	L> LE	ENGTH (PE: (PE: (PE: (PE: (PE: (PE: (PE: (PE:	H: 16 DNA: CSM: CSM: CSM: CEY: CON: CON: CON: Attc Phe attt Ile 20 gct Ala cga Arg gal Glu cat His	Homo CDS (1) 13 cet Pro 5 ggc Gly cgt Arg atc Ile cat His	cgc Arg tac Tyr gcc Ala cat His ggc Gly 70 ttc	aag Lys aca Thr ttc Phe gat Asp 55 gat Asp gcc Ala	ccc Pro cct Pro caa Gln 40 gga Gly gga Gly	gat Asp 25 gtc Val gag Glu tac Tyr ggc Gly acc	Trp 10 ctg Leu tgg Trp gca Ala ccc Pro act Thr 90 ttg	gac Asp agc Ser gac Asp ttt Phe 75 ggt Gly	cca Pro gat Asp atc Ile 60 gac Asp	Asn gag Glu gtg Val 45 atg Met ggt Gly ggg Gly	Gln aca Thr 30 acc Thr atc Ile aag Lys gga Gly caa	Ile 15 gtg Val cca Pro aac Asn gac Asp gac Asp ggac	gat Asp ctg Leu ttt Phe gga Gly 80 tcc ser gtc	96 144 192 240

						53											54	
											-	con	tin	ued				
Arg	Val	Lys 115	Tyr	Gly	Asn	Ala	Asp 120	Gly	Glu	Tyr	Cys	Lys 125	Phe	Pro	Phe			
							aac Asn									432		
							acc Thr									480		
_				_			gaa Glu	_	_			_				528		
_	_		_		_	_	ttt Phe			_		_				576		
							ggc Gly 200									624		
							cgc Arg									672		
		_	_			_	ggt Gly				_		_		_	720		
							ggc Gly									768		
							atg Met									816		
							ttc Phe 280									864		
							ttt Phe									912		
							atg Met									960		
							gac Asp									1008		
	_			_		_	ctt Leu								_	1056		
							tgc 360									1104		
							atc Ile									1152		

						Asn									102
						acc Thr									480
						gaa Glu									528
						ttt Phe									576
						ggc Gly 200									624
						cgc Arg									672
						ggt Gly									720
						ggc Gly									768
						atg Met									816
						ttc Phe 280									864
						ttt Phe									912
		_		_	_	atg Met	_							_	960
						gac Asp									1008
						ctt Leu									1056
		_				tgc 360		_	_		-		_		1104
	_	_	_			atc Ile				_	_				1152
			 _		_	gac Asp	_		_			_	_		1200
-						ccg Pro	-	_		-		_			1248
-		_	 	_	_	gtg Val			_			_			1296

												COII	CIII	uea		
atc t Ile T	yr															1344
agc c Ser L																1392
tgg a Trp S 465																1440
aga t Arg T					_	_		_	_					_		1488
atc g Ile A		_	_			_			-		_	-	-	_		1536
gac c Asp L	eu															1584
ctg a Leu L 5																1632
aaa t Lys S 545																1653
<210 > <211 > <212 > <212 > <213 >	· LE	NGTH PE :	1: 5! PRT	51	sa]	piens	s									
<400>	SE	QUEN	ICE :	14												
Tyr A	sn	Phe	Phe	Pro 5	Arg	Lys	Pro	Lys	Trp 10	Asp	ГÀа	Asn	Gln	Ile 15	Thr	
Tyr A	rg	Ile	Ile 20	Gly	Tyr	Thr	Pro	Asp 25	Leu	Asp	Pro	Glu	Thr 30	Val	Asp	
Asp A		Phe 35	Ala	Arg	Ala	Phe	Gln 40	Val	Trp	Ser	Asp	Val 45	Thr	Pro	Leu	
Arg P	he 10	Ser	Arg	Ile	His	Asp 55	Gly	Glu	Ala	Asp	Ile 60	Met	Ile	Asn	Phe	
Gly A	rg	Trp	Glu	His	Gly 70	Asp	Gly	Tyr	Pro	Phe 75	Asp	Gly	Lys	Asp	Gly 80	
Leu L	eu	Ala	His	Ala 85	Phe	Ala	Pro	Gly	Thr 90	Gly	Val	Gly	Gly	Asp 95	Ser	
His P	he	Asp	Asp 100	Asp	Glu	Leu	Trp	Thr	Leu	Gly	Glu	Gly	Gln 110	Val	Val	
Arg V		Lys 115	Tyr	Gly	Asn	Ala	Asp 120	Gly	Glu	Tyr	Cys	Lys 125	Phe	Pro	Phe	
Leu P	he .30	Asn	Gly	ГÀа	Glu	Tyr 135	Asn	Ser	Сув	Thr	Asp 140	Thr	Gly	Arg	Ser	
Asp G 145	ly	Phe	Leu	Trp	Cys 150	Ser	Thr	Thr	Tyr	Asn 155	Phe	Glu	Lys	Asp	Gly 160	
Lys T	yr	Gly	Phe	Cys	Pro	His	Glu	Ala	Leu 170	Phe	Thr	Met	Gly	Gly 175	Asn	
Ala G	lu	Gly	Gln 180	Pro	Cys	Lys	Phe	Pro 185	Phe	Arg	Phe	Gln	Gly 190	Thr	Ser	
Tyr A		Ser 195	Сув	Thr	Thr	Glu	Gly 200	Arg	Thr	Asp	Gly	Tyr 205	Arg	Trp	СЛа	

-continued

Gly Thr Thr Glu Asp Tyr Asp Arg Asp Lys Lys Tyr Gly Phe Cys I 210 215 220	Pro
Glu Thr Ala Met Ser Thr Val Gly Gly Asn Ser Glu Gly Ala Pro C 225 230 235	Cys 240
Val Phe Pro Phe Thr Phe Leu Gly Asn Lys Tyr Glu Ser Cys Thr S	Ser
Ala Gly Arg Ser Asp Gly Lys Met Trp Cys Ala Thr Thr Ala Asn T	Гуr
Asp Asp Asp Arg Lys Trp Gly Phe Cys Pro Asp Gln Gly Tyr Ser I 275 280 285	Leu
Phe Leu Val Ala Ala His Glu Phe Gly His Ala Met Gly Leu Glu F 290 295 300	His
Ser Gln Asp Pro Gly Ala Leu Met Ala Pro Ile Tyr Thr Tyr Thr I 305 310 315	З20
Asn Phe Arg Leu Ser Gln Asp Asp Ile Lys Gly Ile Gln Glu Leu 1 325 330 335	Гуr
Gly Ala Ser Pro Asp Ile Asp Leu Gly Thr Gly Pro Thr Pro Thr I 340 345 350	Leu
Gly Pro Val Thr Pro Glu Ile Cys Lys Gln Asp Ile Val Phe Asp C 355 360 365	Gly
Ile Ala Gln Ile Arg Gly Glu Ile Phe Phe Phe Lys Asp Arg Phe I 370 375 380	Ile
Trp Arg Thr Val Thr Pro Arg Asp Lys Pro Met Gly Pro Leu Leu V 385 390 395	/al 100
Ala Thr Phe Trp Pro Glu Leu Pro Glu Lys Ile Asp Ala Val Tyr 0 405 410 415	Glu
Ala Pro Gln Glu Glu Lys Ala Val Phe Phe Ala Gly Asn Glu Tyr 7 420 425 430	Гrр
Ile Tyr Ser Ala Ser Thr Leu Glu Arg Gly Tyr Pro Lys Pro Leu 7 435 440 445	Thr
Ser Leu Gly Leu Pro Pro Asp Val Gln Arg Val Asp Ala Ala Phe A 450 455 460	Asn
Trp Ser Lys Asn Lys Lys Thr Tyr Ile Phe Ala Gly Asp Lys Phe 7465 470 475	Ггр 180
Arg Tyr Asn Glu Val Lys Lys Lys Met Asp Pro Gly Phe Pro Lys I 485 490 495	Leu
Ile Ala Asp Ala Trp Asn Ala Ile Pro Asp Asn Leu Asp Ala Val V 500 505 510	/al
Asp Leu Gln Gly Gly His Ser Tyr Phe Phe Lys Gly Ala Tyr 7 515 520 525	Гуr
Leu Lys Leu Glu Asn Gln Ser Leu Lys Ser Val Lys Phe Gly Ser I 530 540	Ile
Lys Ser Asp Trp Leu Gly Cys 545 550	
<pre><210> SEQ ID NO 15 <211> LENGTH: 87 <212> TYPE: DNA <213> ORGANISM: Homo sapiens <220> FEATURE: <221> NAME/KEY: CDS <222> LOCATION: (1)(87) <400> SEQUENCE: 15 atg gag gcg cta atg gcc cgg ggc gcg ctc acg ggt ccc ctg agg gcg</pre>	gcg
Met Glu Ala Leu Met Ala Arg Gly Ala Leu Thr Gly Pro Leu Arg A	Ala

48

								-	con	tin [.]	ued		
1	5					10					15		
ctc tgt ctc ctc Leu Cys Leu Leu 20		_	_	_	_		_	_	_				87
<210> SEQ ID NO <211> LENGTH: 2 <212> TYPE: PRT <213> ORGANISM:	29 [o sar	piens	3									
<400> SEQUENCE:	: 16												
Met Glu Ala Leu 1	Met 5	Ala	Arg	Gly	Ala	Leu 10	Thr	Gly	Pro	Leu	Arg 15	Ala	
Leu Cys Leu Leu 20	ı Gly	Cys	Leu	Leu	Ser 25	His	Ala	Ala	Ala				
<pre><210> SEQ ID NO <211> LENGTH: 1 <212> TYPE: DNA <213> ORGANISM: <220> FEATURE: <221> NAME/KEY: <222> LOCATION:</pre>	L893 A : Homo : CDS			₹									
<400> SEQUENCE:	: 17												
gcg ccg tcg ccc Ala Pro Ser Pro 1													48
gac aaa gag tto Asp Lys Glu Leu 20													96
aag gag agc tgc Lys Glu Ser Cys 35		_			_	_	_			_	_	_	144
cag aag ttc ttt Gln Lys Phe Phe 50													192
atc gag acc atg Ile Glu Thr Met 65		_		_	_				_		_		240
tac aac ttc ttc Tyr Asn Phe Phe		_	_		_		-	_		_			288
tac agg atc att Tyr Arg Ile Ile 100	e Gly												336
gat gcc ttt gct Asp Ala Phe Ala 115	_	_			_		_	_				_	384
cgg ttt tct cga Arg Phe Ser Arg 130													432
ggc cgc tgg gag Gly Arg Trp Glu 145			_					_		_	_		480
ctc ctg gct cat Leu Leu Ala His	_		_					-			_		528
cat ttt gat gad His Phe Asp Asp 180	Asp					_		-				_	576

1		
	-continued	

													tını	uea —			
	gtg Val															624	
	ttc Phe 210															672	
	ggc Gly															720	
	tac Tyr															768	
	gaa Glu															816	
	gac Asp															864	
	acc Thr 290															912	
	acc Thr															960	
	ttc Phe															1008	
_	ggc Gly	_	_	_		_	_		_				_			1056	
	gac Asp															1104	
	ctc Leu 370															1152	
	caa Gln															1200	
	ttc Phe															1248	
	gcc Ala			_		_									_	1296	
	cct Pro	_					_		_	_		_		_		1344	
	gct Ala 450	_		_							_	_				1392	
	cgg Arg			_		_	-	_		_			_	_		1440	
-	aca Thr						_	_	_		_		_			1488	
-	cca Pro	_			_	-				-			-			1536	

	tac Tyr															1584
	ctg Leu 530															1632
	agc Ser															1680
	tac Tyr															1728
	gca Ala															1776
	ctg Leu															1824
_	aag Lys 610	_				_	_	_	_		_			_		1872
	tcc Ser	_				_										1893
	3 > OF O > SE				sa]	pien	3									
<212	L> LE 2> T? 3> OF	PE:	PRT		o saj	pien	5									
	Pro			Ile	Ile	Lys	Phe	Pro	Gly	Asp	Val	Ala	Pro	Lys	Thr	
1 Asp	Lys	Glu	Leu	5 Ala	Val	Gln	Tyr	Leu	10 Asn	Thr	Phe	Tyr	Gly	15 Cys	Pro	
Lys	Glu	Ser	20 Cys	Asn	Leu	Phe	Val	25 Leu	Lys	Asp	Thr	Leu	30 Lys	Lys	Met	
		35					40					45				
GIII	Lys 50	PHE	PHE	GIY	ьец	55	GIII	1111	GIY	Asp	60	Asp	GIII	ASII	1111	
Ile 65	Glu	Thr	Met	Arg	Lys 70	Pro	Arg	Cya	Gly	Asn 75	Pro	Asp	Val	Ala	Asn 80	
Tyr	Asn	Phe	Phe	Pro 85	Arg	Lys	Pro	Lys	Trp 90	Asp	ГÀа	Asn	Gln	Ile 95	Thr	
Tyr	Arg	Ile	Ile 100	Gly	Tyr	Thr	Pro	Asp 105	Leu	Asp	Pro	Glu	Thr 110	Val	Asp	
Asp	Ala	Phe 115	Ala	Arg	Ala	Phe	Gln 120	Val	Trp	Ser	Asp	Val 125	Thr	Pro	Leu	
Arg	Phe 130	Ser	Arg	Ile	His	Asp 135	Gly	Glu	Ala	Asp	Ile 140	Met	Ile	Asn	Phe	
Gly 145	Arg	Trp	Glu	His	Gly 150	Asp	Gly	Tyr	Pro	Phe 155	Asp	Gly	Lys	Asp	Gly 160	
Leu	Leu	Ala	His	Ala 165	Phe	Ala	Pro	Gly	Thr 170	Gly	Val	Gly	Gly	Asp 175	Ser	
His	Phe	Asp	Asp 180	Asp	Glu	Leu	Trp	Thr 185	Leu	Gly	Glu	Gly	Gln 190	Val	Val	
Arg	Val	Lys 195	Tyr	Gly	Asn	Ala	Asp 200	Gly	Glu	Tyr	Сув	Lys 205	Phe	Pro	Phe	

66

_															
Leu	Phe 210	Asn	Gly	ГÀа	Glu	Tyr 215	Asn	Ser	Cys	Thr	Asp 220	Thr	Gly	Arg	Ser
Asp 225	Gly	Phe	Leu	Trp	Cys 230	Ser	Thr	Thr	Tyr	Asn 235	Phe	Glu	Lys	Asp	Gly 240
Lys	Tyr	Gly	Phe	Сув 245	Pro	His	Glu	Ala	Leu 250	Phe	Thr	Met	Gly	Gly 255	Asn
Ala	Glu	Gly	Gln 260	Pro	CAa	Lys	Phe	Pro 265	Phe	Arg	Phe	Gln	Gly 270	Thr	Ser
Tyr	Asp	Ser 275	Сув	Thr	Thr	Glu	Gly 280	Arg	Thr	Asp	Gly	Tyr 285	Arg	Trp	Cys
Gly	Thr 290	Thr	Glu	Asp	Tyr	Asp 295	Arg	Asp	ГЛа	Lys	Tyr 300	Gly	Phe	CÀa	Pro
Glu 305	Thr	Ala	Met	Ser	Thr 310	Val	Gly	Gly	Asn	Ser 315	Glu	Gly	Ala	Pro	Сув 320
Val	Phe	Pro	Phe	Thr 325	Phe	Leu	Gly	Asn	330	Tyr	Glu	Ser	Cys	Thr 335	Ser
Ala	Gly	Arg	Ser 340	Asp	Gly	Lys	Met	Trp 345	Сув	Ala	Thr	Thr	Ala 350	Asn	Tyr
Asp	Asp	Asp 355	Arg	ГÀв	Trp	Gly	Phe 360	Cys	Pro	Asp	Gln	Gly 365	Tyr	Ser	Leu
Phe	Leu 370	Val	Ala	Ala	His	Glu 375	Phe	Gly	His	Ala	Met 380	Gly	Leu	Glu	His
Ser 385	Gln	Asp	Pro	Gly	Ala 390	Leu	Met	Ala	Pro	Ile 395	Tyr	Thr	Tyr	Thr	Lys 400
Asn	Phe	Arg	Leu	Ser 405	Gln	Asp	Asp	Ile	Lys 410	Gly	Ile	Gln	Glu	Leu 415	Tyr
Gly	Ala	Ser	Pro 420	Asp	Ile	Asp	Leu	Gly 425	Thr	Gly	Pro	Thr	Pro 430	Thr	Leu
Gly	Pro	Val 435	Thr	Pro	Glu	Ile	Cys 440	ГÀз	Gln	Asp	Ile	Val 445	Phe	Asp	Gly
Ile	Ala 450	Gln	Ile	Arg	Gly	Glu 455	Ile	Phe	Phe	Phe	Lys 460	Asp	Arg	Phe	Ile
Trp 465	Arg	Thr	Val	Thr	Pro 470	Arg	Asp	Lys	Pro	Met 475	Gly	Pro	Leu	Leu	Val 480
Ala	Thr	Phe	Trp	Pro 485	Glu	Leu	Pro	Glu	Lys 490	Ile	Asp	Ala	Val	Tyr 495	Glu
Ala	Pro	Gln	Glu 500	Glu	Lys	Ala	Val	Phe 505	Phe	Ala	Gly	Asn	Glu 510	Tyr	Trp
Ile	Tyr	Ser 515	Ala	Ser	Thr	Leu	Glu 520	Arg	Gly	Tyr	Pro	Lуз 525	Pro	Leu	Thr
Ser	Leu 530	Gly	Leu	Pro	Pro	Asp 535	Val	Gln	Arg	Val	Asp 540	Ala	Ala	Phe	Asn
Trp 545	Ser	Lys	Asn	ГÀа	Lys 550	Thr	Tyr	Ile	Phe	Ala 555	Gly	Asp	Lys	Phe	Trp 560
Arg	Tyr	Asn	Glu	Val 565	ГÀа	Lys	Lys	Met	Asp 570	Pro	Gly	Phe	Pro	Lys 575	Leu
Ile	Ala	Asp	Ala 580	Trp	Asn	Ala	Ile	Pro 585	Asp	Asn	Leu	Asp	Ala 590	Val	Val
Asp	Leu	Gln 595	Gly	Gly	Gly	His	Ser 600	Tyr	Phe	Phe	Lys	Gly 605	Ala	Tyr	Tyr
Leu	Lys 610	Leu	Glu	Asn	Gln	Ser 615	Leu	Lys	Ser	Val	Lys 620	Phe	Gly	Ser	Ile

Lys Ser Asp Trp Leu Gly Cys													
Lys Ser Asp Trp 625	Leu Gly Cys 630												
<pre><210> SEQ ID NO <211> LENGTH: 2 <212> TYPE: DNA <213> ORGANISM: <220> FEATURE: <222> OTHER INF <220> FEATURE: <221> NAME/KEY: <222> LOCATION:</pre>	490 Artificial ORMATION: fusic	on protein											
<400> SEQUENCE:	19												
		gc gcg ctc acg ggt Y Ala Leu Thr Gly 10											
		g agc cac gcc gcc u Ser His Ala Ala 25											
		at gtc gcc ccc aaa pp Val Ala Pro Lys)											
		cc ttc tat ggc tgc nr Phe Tyr Gly Cys 60											
		ac aca cta aag aag sp Thr Leu Lys Lys 75											
		at ctt gac cag aat pp Leu Asp Gln Asn 90											
		ac cca gat gtg gcc sn Pro Asp Val Ala 105											
		ac aag aac cag atc sp Lys Asn Gln Ile 20											
		ac cca gag aca gtg sp Pro Glu Thr Val 140											
		gc gat gtg acc cca er Asp Val Thr Pro 155											
		ac atc atg atc aac sp Ile Met Ile Asn 170											
		t gac ggt aag gac ne Asp Gly Lys Asp 185											
-		gt gtt ggg gga gac .y Val Gly Gly Asp 00	=										
		ga gaa ggc caa gtg .y Glu Gly Gln Val 220											
		ac tgc aag ttc ccc or Cys Lys Phe Pro 235	_										
		et gat acc ggc cgc nr Asp Thr Gly Arg 250											

ctc	tga	tqc	tcc	acc	acc	tac	aac	ttt	gaq	aaq	gat	gqc	aaq	tac	gqc	816
		_			Thr					_	_		_			
					gcc Ala											864
_		_	_		cca Pro		_		_					_	_	912
					cgc Arg 310											960
					gac Asp											1008
					gly ggg											1056
			_		aac Asn				_	_		_	_		_	1104
					tgg Trp											1152
					tgc Cys 390											1200
_	_				ggc Gly		_	_		_					_	1248
					gca Ala											1296
					atc Ile											1344
					ggc Gly											1392
					aaa Lys 470											1440
					ttc Phe											1488
					aag Lys											1536
					gaa Glu											1584
		_	-		ttc Phe		-			-						1632
					cga Arg 550											1680
					cag Gln											1728

													COII	CIII	ucu			
-					565					570					575			
										gac Asp							1776	
										ttt Phe							1824	
				_			_		_	gat Asp	-	_		_	_	_	1872	
C										ggt Gly							1920	
										ttt Phe 650							1968	
										gag Glu							2016	
										acc Thr							2064	
										ccg Pro							2112	
ľ										tgc Cys							2160	
										aag Lys 730							2208	
										aaa Lys							2256	
										gag Glu							2304	
										cag Gln							2352	
C										gcc Ala							2400	
										tgt Cys 810							2448	
										gly aaa							2490	
<	<210> SEQ ID NO 20																	

<210> SEQ ID NO 20 <211> LENGTH: 830

<400> SEQUENCE: 20

Met Glu Ala Leu Met Ala Arg Gly Ala Leu Thr Gly Pro Leu Arg Ala 1 $$ 5 $$ 10 $$ 15

<212> TYPE: PRT <213> ORGANISM: Artificial

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic Construct

Leu	Cys	Leu	Leu 20	Gly	CAa	Leu	Leu	Ser 25	His	Ala	Ala	Ala	Ala 30	Pro	Ser
Pro	Ile	Ile 35	Lys	Phe	Pro	Gly	Asp 40	Val	Ala	Pro	Lys	Thr 45	Asp	Lys	Glu
Leu	Ala 50	Val	Gln	Tyr	Leu	Asn 55	Thr	Phe	Tyr	Gly	Сув 60	Pro	Lys	Glu	Ser
Cys 65	Asn	Leu	Phe	Val	Leu 70	Lys	Asp	Thr	Leu	Lys 75	ГÀа	Met	Gln	Lys	Phe 80
Phe	Gly	Leu	Pro	Gln 85	Thr	Gly	Asp	Leu	Asp 90	Gln	Asn	Thr	Ile	Glu 95	Thr
Met	Arg	Lys	Pro 100	Arg	Cys	Gly	Asn	Pro 105	Asp	Val	Ala	Asn	Tyr 110	Asn	Phe
Phe	Pro	Arg 115	Lys	Pro	Lys	Trp	Asp 120	Lys	Asn	Gln	Ile	Thr 125	Tyr	Arg	Ile
Ile	Gly 130	Tyr	Thr	Pro	Asp	Leu 135	Asp	Pro	Glu	Thr	Val 140	Asp	Asp	Ala	Phe
Ala 145	Arg	Ala	Phe	Gln	Val 150	Trp	Ser	Asp	Val	Thr 155	Pro	Leu	Arg	Phe	Ser 160
Arg	Ile	His	Asp	Gly 165	Glu	Ala	Asp	Ile	Met 170	Ile	Asn	Phe	Gly	Arg 175	Trp
Glu	His	Gly	Asp 180	Gly	Tyr	Pro	Phe	Asp 185	Gly	Lys	Asp	Gly	Leu 190	Leu	Ala
His	Ala	Phe 195	Ala	Pro	Gly	Thr	Gly 200	Val	Gly	Gly	Asp	Ser 205	His	Phe	Asp
Asp	Asp 210	Glu	Leu	Trp	Thr	Leu 215	Gly	Glu	Gly	Gln	Val 220	Val	Arg	Val	Lys
Tyr 225	Gly	Asn	Ala	Asp	Gly 230	Glu	Tyr	Cys	Lys	Phe 235	Pro	Phe	Leu	Phe	Asn 240
Gly	Lys	Glu	Tyr	Asn 245	Ser	Càa	Thr	Asp	Thr 250	Gly	Arg	Ser	Asp	Gly 255	Phe
Leu	Trp	СЛа	Ser 260	Thr	Thr	Tyr	Asn	Phe 265	Glu	Lys	Asp	Gly	Lys 270	Tyr	Gly
Phe	Сув	Pro 275	His	Glu	Ala	Leu	Phe 280	Thr	Met	Gly	Gly	Asn 285	Ala	Glu	Gly
Gln	Pro 290	СЛа	ГÀа	Phe	Pro	Phe 295	Arg	Phe	Gln	Gly	Thr 300	Ser	Tyr	Asp	Ser
305 Cys	Thr	Thr	Glu	Gly	Arg 310	Thr	Asp	Gly	Tyr	Arg 315	Trp	CAa	Gly	Thr	Thr 320
Glu	Asp	Tyr	Asp	Arg 325	Asp	ГЛа	Lys	Tyr	Gly 330	Phe	CÀa	Pro	Glu	Thr 335	Ala
Met	Ser	Thr	Val 340	Gly	Gly	Asn	Ser	Glu 345	Gly	Ala	Pro	CAa	Val 350	Phe	Pro
Phe	Thr	Phe 355	Leu	Gly	Asn	ГÀв	Tyr 360	Glu	Ser	Cys	Thr	Ser 365	Ala	Gly	Arg
Ser	Asp 370	Gly	Lys	Met	Trp	Cys 375	Ala	Thr	Thr	Ala	Asn 380	Tyr	Asp	Asp	Asp
Arg 385	Lys	Trp	Gly	Phe	Сув 390	Pro	Asp	Gln	Gly	Tyr 395	Ser	Leu	Phe	Leu	Val 400
Ala	Ala	His	Glu	Phe 405	Gly	His	Ala	Met	Gly 410	Leu	Glu	His	Ser	Gln 415	Asp
Pro	Gly	Ala	Leu 420	Met	Ala	Pro	Ile	Tyr 425	Thr	Tyr	Thr	Lys	Asn 430	Phe	Arg
Leu	Ser	Gln	Asp	Asp	Ile	Lys	Gly	Ile	Gln	Glu	Leu	Tyr	Gly	Ala	Ser

Pro A	Asp 450	Tle													
Thr E			Asp	Leu	Gly	Thr 455	Gly	Pro	Thr	Pro	Thr 460	Leu	Gly	Pro	Val
465	Pro	Glu	Ile	Cys	Lys 470	Gln	Asp	Ile	Val	Phe 475	Asp	Gly	Ile	Ala	Gln 480
Ile A	Arg	Gly	Glu	Ile 485	Phe	Phe	Phe	Lys	Asp 490	Arg	Phe	Ile	Trp	Arg 495	Thr
Val 1	Thr	Pro	Arg 500	Asp	Lys	Pro	Met	Gly 505	Pro	Leu	Leu	Val	Ala 510	Thr	Phe
Trp I	Pro	Glu 515	Leu	Pro	Glu	Lys	Ile 520	Asp	Ala	Val	Tyr	Glu 525	Ala	Pro	Gln
Glu (Glu 530	Lys	Ala	Val	Phe	Phe 535	Ala	Gly	Asn	Glu	Tyr 540	Trp	Ile	Tyr	Ser
Ala S 545	Ser	Thr	Leu	Glu	Arg 550	Gly	Tyr	Pro	ГЛа	Pro 555	Leu	Thr	Ser	Leu	Gly 560
Leu I	Pro	Pro	Asp	Val 565	Gln	Arg	Val	Asp	Ala 570	Ala	Phe	Asn	Trp	Ser 575	Lys
Asn I	Lys	Lys	Thr 580	Tyr	Ile	Phe	Ala	Gly 585	Asp	Lys	Phe	Trp	Arg 590	Tyr	Asn
Glu \	Val	Lys	Lys	Lys	Met	Asp	Pro 600	Gly	Phe	Pro	ГÀа	Leu 605	Ile	Ala	Asp
Ala 1	Trp 610	Asn	Ala	Ile	Pro	Asp 615	Asn	Leu	Asp	Ala	Val 620	Val	Asp	Leu	Gln
Gly (625	Gly	Gly	His	Ser	Tyr 630	Phe	Phe	ГЛа	Gly	Ala 635	Tyr	Tyr	Leu	Lys	Leu 640
Glu A	Asn	Gln	Ser	Leu 645	Lys	Ser	Val	ГЛа	Phe 650	Gly	Ser	Ile	Lys	Ser 655	Asp
Trp I	Leu	Gly	Cys 660	Glu	Phe	Lys	Pro	Thr 665	Glu	Asn	Asn	Glu	Asp 670	Phe	Asn
Ile V	Val	Ala 675	Val	Ala	Ser	Asn	Phe 680	Ala	Thr	Thr	Asp	Leu 685	Asp	Ala	Asp
Arg (Gly 690	Lys	Leu	Pro	Gly	Lys 695	Lys	Leu	Pro	Leu	Glu 700	Val	Leu	Lys	Glu
Met (Glu	Ala	Asn	Ala	Arg 710	Lys	Ala	Gly	Сла	Thr 715	Arg	Gly	Cys	Leu	Ile 720
Cys I	Leu	Ser	His	Ile 725	Lys	Cys	Thr	Pro	Lys 730	Met	ГÀЗ	ГÀЗ	Phe	Ile 735	Pro
Gly A	Arg	Cys	His 740	Thr	Tyr	Glu	Gly	Asp 745	Lys	Glu	Ser	Ala	Gln 750	Gly	Gly
Ile (Gly	Glu 755	Ala	Ile	Val	Asp	Ile 760	Pro	Glu	Ile	Pro	Gly 765	Phe	Lys	Asp
Leu (Glu 770	Pro	Met	Glu	Gln	Phe 775	Ile	Ala	Gln	Val	Asp 780	Leu	Cys	Val	Asp
Cys 7	Thr	Thr	Gly	CAa	Leu 790	Lys	Gly	Leu	Ala	Asn 795	Val	Gln	Cys	Ser	Asp
Leu I	Leu	Lys	Lys	Trp 805	Leu	Pro	Gln	Arg	Cys 810	Ala	Thr	Phe	Ala	Ser 815	Lys
Ile (Gln	Gly	Gln 820	Val	Asp	Lys	Ile	Lys 825	Gly	Ala	Gly	Gly	Asp 830		

<210> SEQ ID NO 21 <211> LENGTH: 36 <212> TYPE: DNA <213> ORGANISM: Artificial

	-continued
<pre><220> FEATURE: <223> OTHER INFORMATION: primer</pre>	
<400> SEQUENCE: 21	
ctcggatcca gccaccatgg ccctgtggat gcgc	cct 36
<pre><210> SEQ ID NO 22 <211> LENGTH: 30 <212> TYPE: DNA <213> ORGANISM: Artificial <220> FEATURE: <223> OTHER INFORMATION: primer</pre>	
<400> SEQUENCE: 22	
cttgaattcg ttgcagtagt tctccagctg	30
<pre><210> SEQ ID NO 23 <211> LENGTH: 35 <212> TYPE: DNA <213> ORGANISM: Artificial <220> FEATURE: <223> OTHER INFORMATION: primer</pre>	
<pre><400> SEQUENCE: 23 ggcaagetta gccaccatgg aggcgctaat ggcc</pre>	cc 35
<pre><210> SEQ ID NO 24 <211> LENGTH: 28 <212> TYPE: DNA <213> ORGANISM: Artificial <220> FEATURE: <223> OTHER INFORMATION: primer</pre>	
<400> SEQUENCE: 24	
ggcgaattcg cagcctagcc agtcggat	28

The invention claimed is:

- 1. A method of screening a substance regulating insulin 40 secretion from a cell, which comprises:
 - contacting a cell transformed with a polynucleotide encoding a fusion protein of preproinsulin and a luciferase with a test substance, and
 - detecting luminescence of the luciferase to measure a level 45 of insulin secretion,
 - wherein the luciferase is Gaussia luciferase, and
 - wherein *Gaussia* luciferase is a protein of any one of (a) through (c) below:
 - (a) a protein comprising the amino acid sequence of SEQ 50 ID NO: 8;
 - (b) a protein consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of 1 to 10 amino acid residues in the amino acid sequence of SEQ ID NO: 8, and having substantially the 55 same activity as a protein consisting of the amino acid sequence of SEQ ID NO: 8;
 - (c) a protein consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 8, and having substantially the same activity as the 60 protein consisting of the amino acid sequence of SEQ ID NO: 8; and,
 - wherein the preproinsulin consists of a signal peptide of preproinsulin and the polypeptide of any one of (i) through (k) below:
 - (i) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 4;

- (j) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of 1 to 10 amino acid residues in the amino acid sequence of SEQ ID NO: 4, and having an insulin activity;
- (k) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 4, and having an insulin activity; and,
- wherein the signal peptide of preproinsulin is a polypeptide consisting of the amino acid sequence of SEQ ID NO: 6.
- 2. The screening method according to claim 1, wherein the any one of (e) through (g) below:
 - (e) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 10;
 - (f) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of 1 to 10 amino acid residues in the amino acid sequence of SEQ ID NO: 10, and having an insulin activity;
 - (g) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 10, and having an insulin activity; and,
 - (h) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 9

- wherein the high stringent conditions are $5\times SSC$, $5\times Denhardt's$ solution, 0.5% (w/v) SDS, 50% (v/v) formamide and 50° C., and having an insulin activity.
- 3. The screening method according to claim 1, wherein the fusion any one of (m) through (o) below:
 - (m) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 12;
 - (n) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of 1 to 10 amino acid residues in the amino acid sequence of SEQ ID NO: 12, and having an insulin activity;
 - (o) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 12, and having an insulin activity; and,
 - (p) a polypeptide comprising a polypeptide consisting of an amino acid sequence encoded by a polynucleotide that hybridizes under high stringent conditions to a polynucleotide consisting of a nucleotide sequence complementary to the nucleotide sequence of SEQ ID NO: 11 wherein the high stringent conditions are 5xSSC, 5xDenhardt's solution, 0.5% (w/v) SDS, 50% (v/v) formamide and 50° C., and having an insulin activity.
- **4**. The screening method according to claim **1**, which further comprises the step of detecting luminescence using a CCD camera or a photon counting camera.
- 5. A method of screening a substance regulating insulin secretion from a cell, which comprises:
 - contacting a cell transformed with a polynucleotide encoding a fusion protein of preproinsulin and a luciferase with a test substance, and
 - detecting luminescence of the luciferase to measure a level of insulin secretion,

80

- wherein the luciferase is *Gaussia* luciferase, and wherein *Gaussia* luciferase is a protein of any one of (a) through (c) below:
- (a) a protein consisting of the amino acid sequence of SEQ ID NO: 8;
- (b) a protein consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of 1 to 5 amino acid residues in the amino acid sequence of SEQ ID NO: 8, and having substantially the same activity as a protein consisting of the amino acid sequence of SEQ ID NO: 8;
- (c) a protein consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 8, and having substantially the same activity as the protein consisting of the amino acid sequence of SEQ ID NO: 8; and.
- wherein the preproinsulin consists of a signal peptide of preproinsulin and the polypeptide of any one of (i) through (k) below:
- (i) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 4;
- (j) a polypeptide comprising a polypeptide consisting of an amino acid sequence comprising deletion, substitution, insertion and/or addition of 1 to 10 amino acid residues in the amino acid sequence of SEQ ID NO: 4, and having an insulin activity;
- (k) a polypeptide comprising a polypeptide consisting of an amino acid sequence having at least 90% identity to the amino acid sequence of SEQ ID NO: 4, and having an insulin activity; and,
- wherein the signal peptide of preproinsulin is a polypeptide consisting of the amino acid sequence of SEQ ID NO: 6.

* * * * *